APPLICATION FOR UNITED STATES PATENT

INVENTOR(S):   Robert G. Ulrich
Mark A. Olson
Sina Bavari

INVENTION: Bacterial Superantigen Vaccines

SPECIFICATION

SAP 9/94

TITLE OF THE INVENTION

Bacterial Superantigen Vaccines
by
5   Robert G. Ulrich
Mark A. Olson
Sina Bavari

INTRODUCTION

10

Staphylococcal enterotoxins (SEs) A through E are the most commom cause of food poisoning [Bergdoll, M.S.(1983) In Easom CSF, Aslam C., eds. Staphylococci and staphylococcal infections. London: Academic Press, 15   pp 559-598] and are associated with several serious diseases [Schlievert, P.M. (1993) *J. Infect. Dis.* **167**: 997-1002; Ulrich *et al.* (1995) *Trends Microbiol.* **3**: 463-468], such as bacterial arthritis [Schwab *et al.* (1993) *J. Immunol.* **150**; 4151-4159; Goldenberg *et* 20   *al.* (1985) *N. Engl. J. Med.* **312**: 764-771], other autoimmune disorders [Psnett, D. N. (1993) *Semin. Immunol.* **5**: 65-72], and toxic shock syndrome [Schlieverst, P.M. (1986) *Lancet* **1**: 1149-1150; Bohach *et al.* (1990) *Crit. Rev. Microbiol.* **17**: 251-272]. The 25   nonenterotoxic staphylococcal superantigen toxic shock syndrome toxin-1 (TSST-1) was first identified as a causative agent of menstrual-associated toxic shock syndrome [Schlievert *et al.* (1981) *J. Infect. Dis.* **143**: 509-516]. Superantigen-producing *Staphylococcus* 30   *aureus* strains are also linked to Kawasaki syndrome, an inflammatory disease of children [Leung *et al.* (1993) *Lancet* **342**: 1385-1388].

The staphylococcal enterotoxins A-E, toxic shock syndrome toxin-1 (TSST-1), and streptococcal pyrogenic

exotoxins A-C are soluble 23-29-kD proteins commonly referred to as bacterial superantigens (SAgs). Bacterial superantigens are ligands for both major histocompatibility complex (MHC) class II molecules,

5 expressed on antigen-presenting cells, and the variable portion of the T cell antigen receptor β chain (TCR Vβ) [Choi *et al.* (1989) *Proc. Natl. Acad. Sci. USA* **86**:8941-8945; Fraser, J.D. (1989) *Nature* **339**:221-223; Marrack *et al.* (1990) *Science* **248**: 705-711; Herman *et*

10 *al.* (1991) *Annu. Rev. Immunol.* **9**: 745-772; Mollick *et al.* (1989) *Science* **244**:817-820].

Each bacterial superantigen has a distinct affinity to a set of TCR Vβ, and coligation of the MHC class II molecule polyclonally stimulates T cells

15 [White *et al.* (1989) *Cell* **56**: 27-35; Kappler *et al.* (1989) *Science* **244**: 811-813; Takimoto *et al.* (1990) *Eur J. Immunol.* **140**: 617-621]. Pathologically elevated levels of cytokines that are produced by activated T cells are the probable cause of toxic

20 shock symptoms [Calson *et al.* (1985) *Cell. Immunol.* **96**: 175-183; Stiles *et al.* (1993) *Infect. Immun.* **61**: 5333-5338]. In addition, susceptibility to lethal gram-negative endotoxin shock is enhanced by several bacterial superantigens [Stiles, *et al., supra*].

25 Although antibodies reactive with superantigens are present at low levels in human sera [Takei *et al.* (1993) *J. Clin. Invest.* **91**: 602-607], boosting antibody titers by specific immunization may be efficacious for patients at risk for toxic shock

30 syndrome and the other disorders of common etiology.

A vaccine approach to controlling bacterial superantigen-associated diseases presents a unique set of challenges. Acute exposure to bacterial superantigens produces T cell anergy, a state of

specific non-responsiveness [Kawabe *et al.* (1991) *Nature* **349**: 245-248], yet T cell help is presumably a requirement for mounting an antibody response.

Presently, the only superantigen vaccines
5   available are chemically inactivated toxoids from different bacteria which have several disadvantages. The chemical inactivation process can be variable for each production lot making the product difficult to characterize. The chemical used for inactivation,
10   (e.g. formaldehyde), is often toxic and does not negate the possibility of reversion of the inactivated superantigen to an active form. In addition, the yields of wild-type toxin from bacterial strains used for making toxoids are often low.

15

SUMMARY OF THE INVENTION

The present invention relates to a vaccine which overcomes the disadvantages of the chemically inactivated toxoids described above. The superantigen
20   vaccine(s) of the present invention is/are designed to protect individuals against the pathologies resulting from exposure to one or several related staphylococcal and streptococcal toxins. The superantigen vaccine is comprised of a purified protein product that is
25   genetically attenuated by DNA methodologies such that superantigen attributes are absent, however the superantigen is effectively recognized by the immune system and an appropriate antibody response is produced.
30   Specifically, the vaccine of the present invention is a product of site-directed mutagenesis of the DNA coding sequences of superantigen toxins resulting in a disruption of binding to both the MHC class II receptor and to the T-cell antigen receptor.
35   A comprehensive study of the relationships of the

superantigen structures of TSST-1, streptococcal pyrogenic exotoxin-A (SPEa), staphylococcal enterotoxin B (SEB), and staphylococcal enterotoxin A, to receptor binding were undertaken to provide insight into the design of the vaccine. From these studies, critical amino acid residues of the toxin responsible for binding the superantigen to the human MHC receptor were defined. Site-directed mutagenesis of the gene encoding the toxin and expression of the new protein product resulted in a superantigen toxin with disrupted binding to the MHC receptors.

Therefore, it is an object of the present invention to provide a superantigen toxin DNA fragment which has been genetically altered such that binding of the encoded altered toxin to the MHC class II or T-cell antigen receptor is disrupted. Such a DNA fragment is useful in the production of a vaccine against superantigen toxin infections.

It is another object of the present invention to provide a superantigen toxin amino acid sequence which has been altered such that the binding of the encoded altered toxin to the MHC class II or T-cell antigen receptor is disrupted. Such a sequence is useful for the production of a superantigen toxin vaccine.

It is another object of the invention to provide a recombinant vector comprising a vector and the DNA fragment described above.

It is a further object of the present invention to provide host cells transformed with the above-described recombinant DNA constructs. Host cells include cells of other prokaryotic species or eukaryotic plant or animal species, including yeasts,

fungi, plant culture, mammalian and nonmammalian cell lines, insect cells and transgenic plants or animals.

It is another object of the present invention to provide a method for producing altered superantigen
5   toxin with disrupted MHC class II and T-cell antigen receptor binding which comprises culturing a host cell under conditions such that a recombinant vector comprising a vector and the DNA fragment described above is expressed and altered superantigen toxin is
10   thereby produced, and isolating superantigen toxin for use as a vaccine against superantigen toxin-associated bacterial infections and as a diagnostic reagent.

It is still another object of the invention to provide a purified altered superantigen toxin useful
15   as a vaccine and as a diagnostic agent.

It is another object of the invention to provide a purified altered superantigen toxin for the therapeutic stimulation of, or other in vivo manipulation of, selective T cell subsets, or ex vivo
20   manipulation of T cells for in vivo therapeutic purposes in mammals. Diseases, such as autoimmunity, wherein T-cell responses of limited diversity (oligoclonal) are evident. Altered superantigens may be used to therapeutically inactivate (induce anergy
25   in) T cells in diseases wherein oligoclonal T-cell responses are evident such as autoimmune diseases, for example. For diseases in which specific T-cell subsets are not active or are anergetic, altered superantigens may be used to therapeutically stimulate
30   these T cells. Such disease include, but are not limited to, infectious diseases and cancers wherein specific subsets of cytotoxic or helper T cells are inactivated or are otherwise unable to respond normally to the antigenic stimulation of the disease
35   moiety.

It is a further object of the present invention to provide an antibody to the above-described altered superantigen toxin for use as a therapeutic agent and as a diagnostic agent.

5   It is yet another object of the invention to provide a superantigen toxin vaccine comprising an altered superantigen toxin effective for the production of antigenic and immunogenic response resulting in the protection of an animal against

10   superantigen toxin infection.

It is a further object of the invention to provide a multivalent superantigen toxin vaccine comprising altered toxins from a variety of streptococcal and staphylococcal toxins effective for

15   the production of antigenic and immunogenic response resulting in the protection of an animal against infection with bacterial superantigen toxin-expressing strains and against other direct or indirect exposures to bacterial superantigen toxins such as might occur

20   by ingestion, inhalation, injection, transdermal or other means.

It is yet another object of the present invention to provide a method for the diagnosis of superantigen toxin-associated bacterial infection comprising the

25   steps of:

(i) contacting a sample from an individual suspected of having a superantigen toxin-associated bacterial infection with antibodies which recognize superantigen toxin using antibodies generated from the

30   altered superantigen toxin; and

(ii) detecting the presence or absence of a superantigen-associated bacterial infection by detecting the presence or absence of a complex formed between superantigen toxin in said sample and

35   antibodies specific therefor.

It is yet another object of the present invention to provide a method for the diagnosis of superantigen bacterial infection comprising the steps of:

(i) contacting a sample from an individual suspected of having the disease with lymphocytes which recognize superantigen toxin produced by said superantigen bacteria or lymphocytes which recognize altered superantigen toxin; and

(ii) detecting the presence or absence of responses of lymphocytes resulting from recognition of superantigen toxin. Responses can be, for example, measured cytokine release, increase of activation markers, mitotic activity, or cell lysis. The lymphocytes responding to the altered superantigen toxins recognize them as recall antigens not as superantigens, therefore the response is an indicator of prior exposure to the specific superantigen. The absence of a response may indicate no prior exposure, a defective immune response or in some cases a manifestation of T-cell anergy. Anergy is defined here as antigen-specific or a generalized non-responsiveness of subsets of T cells.

It is a further object of the present invention to provide a diagnostic kit comprising an antibody against an altered superantigen toxin and ancillary reagents suitable for use in detecting the presence of superantigen toxin in animal tissue or serum.

It is another object of the present invention to provide a detection method for detecting superantigen toxins or antibodies to superantigen toxin in samples, said method comprising employing a biosensor approach. Such methods are known in the art and are described for example in Karlsson *et al.* (1991) *J. Immunol. Methods* **145**, 229-240 and Jonsson *et al.* (1991) *Biotechniques* **11**, 620-627.

It is yet another object of the present invention to provide a therapeutic method for the treatment or amelioration of symptoms of superantigen-associated bacterial infection, said method comprising providing

5 to an individual in need of such treatment an effective amount of sera from individuals immunized with one or more altered superantigen toxins from different bacteria in a pharmaceutically acceptable excipient.

10 It is further another object of the present invention to provide a therapeutic method for the treatment or amelioration of symptoms of superantigen toxin-associated bacterial infection, said method comprising providing to an individual in need of such

15 treatment an effective amount of antibodies against altered superantigen toxins in a pharmaceutically acceptable excipient.

It is another object of the present invention to provide a therapeutic method for the treatment or

20 amelioration of symptoms of bacterial superantigen toxin infection, said method comprising providing to an individual in need of such treatment an effective amount of altered superantigen from a variety of streptococcal and staphylococcal bacteria in order to

25 inhibit adhesion of superantigen bacterial toxin to MHC class II or T-cell receptors by competitive inhibition of these interactions.

It is yet another object of the present invention to provide a therapeutic method for the treatment of

30 diseases that may not be associated directly with superantigen toxins but which result in specific nonresponsiveness of T-cell subsets, said method comprising the administration of altered superantigen toxins, *in vivo* or *ex vivo*, such that T-cell subsets

35 are expanded or stimulated. Diseases which cause

anergy or nonresponsiveness of T-cells include, but are not limited to, infectious diseases.

It is another object of the present invention to provide a therapeutic method for the treatment of

5   diseases associated with expanded or over-stimulated T-cell subsets, such as autoimmunity for example, said method comprising administration of altered superantigen toxin, *in vivo* or *ex vivo*, such that anergy or inactivation of disease associated T-cells

10 is produced. In this case, superantigen mutants can be designed with altered but not attenuated T-cell receptor binding, to cause anergy of only the select (i.e. 1-3) T-cell subsets that are pathologically activated.

15

BRIEF DESCRIPTION OF THE DRAWINGS

These and other features, aspects, and advantages of the present invention will become better understood with reference to the following description and

20 appended claims, and accompanying drawings where:

Figure 1. Staphylococcal and streptococcal superantigen amino acid sequence homologies, compiled with Genetics Computers Group of Univ. of Wisconsin

25 software.

Figure 2. Comparison of mutant SEB and SEA biological activities.

A. SEB mutant HLA-DR1-binding; **B.** SEA mutant HLA-DR1-binding; **C.** T-cell recognition of SEA and SEB

30 mutants. Binding of bacterial superantigens to cell surface DR1 was measured by laser fluorescence-activated flow cytometry. A representative experiment of three performed is shown. The mutants SEA D197N, the homologous SEB D199N, and SEA L11Y had no effect

35 on binding or T-cell recognition, and were used for

controls. Human T-cell proliferation, assessed by
[^{3}H]thymidine incorporation, was measured in response
to SEA (Y64A) or SEB (Y61A) mutants and controls that
retained DR1-binding affinities. Each data point

5 represents the mean of triplicate determinations; SEM
<5%.

Figure 3. Sequence and secondary structural
alignment of bacterial superantigen toxins. Analyses
were performed with the applications PILEUP and

10 PROFILE from the Computer Genetics Group (Madison, WI)
using sequence data obtained from a variety of
sources. Amino acid residue numbering is based on the
SEA sequence.

Figure 4. Detection of TNF-α (a), IL-1α (B), IL-

15 6 (C) and IFN-γ (D) in the serum of mice injected with
SEA (open circles), LPS (open triangles), or SEA plus
LPS (open squares). Values for TNF-α and IL-1α
represent the mean of duplicate samples, with an SEM
of ± 5%. INF-γ and IL-6 values represent the mean of

20 duplicate and triplicate samples, respectively. The
SEMs for IFN-γ and IL-6 readings were ± 5% and ± 10%,
respectively.

Figure 5. Mutant SEA vaccines that have
attenuated major histocompatibility complex class II

25 or T-cell antigen receptor binding do not induce T-
cell anergy. Mice were given three doses of wild type
(WT) SEA or site-specific mutant vaccine, plus
adjuvant. Control animals received adjuvant alone or
were untreated; 2 weeks after final injection, pooled

30 mononuclear cells were collected from spleens of 4
mice from each group. Results are represented as mean
cpm (±SD) of quadruplicate wells incubated with 100
ng/ml WT SEA for 72 h and then pulse-labeled for 12 h
with [^{3}H]thymidine. P<0.0001 (analysis of variance

for repeated measures comparing untreated, adjuvant, Y64A, and Y92A to WT SEA group).

Figure 6.   No superantigen-induced T-cell anergy is exhibited by rhesus monkeys immunized with the

5   vaccine B899445.   Peripheral blood lymphocytes were incubated with titrated concentrations of wild-type superantigens from individual rhesus monkeys (K422 and N103) that were immunized with B899445.   T-cell proliferation was assessed by [^{3}H]thymidine

10   incorporation.   Each data point represents the mean of triplicate determinations; SEM <5%.

Figure 7. Antibody responses of rhesus monkeys immunized with a combined vaccine consisting of B899445 (SEB) and A489270 (SEA).   The antibody levels

15   were measured by ELISA, using plates coated with SEA, SEB or SEC1 as listed.   Monkey G8 is a non-immunized control.   SEM <5% for triplicate measurements.

DETAILED   DESCRIPTION

20   The present invention relates in part to a vaccine against superantigen toxin-associated bacterial diseases.   The superantigen vaccines used in this study were developed by engineering changes in the receptor-binding portions of superantigen toxins

25   to reduce receptor-binding affinities and toxicity while maintaining antigenicity.

Five different superantigen vaccines are described in this application: staphylococcal enterotoxin A, staplylococcal enterotoxin B,

30   staphylococcal enterotoxin C1, toxic-shock syndrome toxin-1, and streptococcal pyrogenic exotoxin-A.   For each of the superantigen toxins above, a comprehensive study of the relationships of the toxin protein structure to receptor binding was undertaken to

35   provide insight into the design of the vaccine.   The

study employed site-directed mutagenesis of toxin and receptor molecules, molecular modeling, protein structure and binding studies. Following these studies, toxins were altered by site-directed

5 mutagenesis to attenuate MHC class II binding and biological activity to an essentially non-specific level. The engineered vaccines were evaluated at each stage of analysis to determine mouse and human T-cell reactivities *in vitro*, serological responses and

10 toxicity in mice and monkeys.

In one embodiment, the present invention relates to an altered superantigen toxin protein having an amino acid sequence which has been altered such that the binding of the toxin to the MHC class II receptor

15 is disrupted.

Comparison of amino acid sequences (**Fig. 1**) suggested that bacterial superantigens fall into groups consisting of (1) SEA, SED and SEE, (2) SEB, staphylococcal enterotoxins C1-C3 (SEC1-3), the

20 streptococcal pyrogenic exotoxins A (SPE-A) and C (SPE-C), (3) TSST-1 and (4) the exfoliative toxins (ETA, ETB) and streptococcal pyrogenic exotoxin B (SPE-B), which are the most distant from the others in sequence. Although not available to the inventor when

25 the inventions were first conceived and proof of principle was obtained, the x-ray crystallographic structures of several bacterial superantigens are now known. Diverse superantigens, such as SEB and TSST-1, appear to have little sequence in common, yet they

30 exhibit homologous protein folds composed largely of β strands [Prasad, G.S. *et al.* (1993) *Biochemistry* **32**, 13761-13766; Acharya, R.K. *et al.* (1994) *Nature* **367**, 94-97; Swaminathan, S. *et al.* (1992) *Nature* **359**, 801-806] within two distinct domains. Differences between

35 the proteins are located primarily in highly variable

regions comprised of several surface loops, such as the disulfide-bonded loop which is absent from TSST-1 and at the amino terminus.

The X-ray crystal structures of SEB and TSST-1
5   complexed with HLA DR1 are known [Kim, J. *et al.* (1994) *Science* **266**, 1870-1874 ; Jardetzky, T.S. *et al.* (1994) *Nature* **368**, 711-718]. The region of HLA DR1 that contacts SEB consists exclusively of α subunit surfaces. The main regions of SEB involved are two
10 conserved sites: a polar pocket derived from three β strands of the β barrel domain and a highly solvent-exposed hydrophobic reverse turn. The polar binding pocket of SEB contains a glutamate and two tyrosines that accommodate Lys39 of the α subunit of HLA DR1,
15 while the hydrophobic region consists of a leucine and flanking residues that make several contacts with the HLA DRα chain. The HLA DR1 binding sites of both TSST-1 and SEB overlap significantly. The hydrophobic binding contacts of other SAg with the HLA DRα chain
20 have been proposed [Ulrich, *et al.* (1995). *Nature*, *Struct. Biol* **2**, 554-560] to be similar to those found in SEB and TSST-1. A motif consisting of a leucine in a reverse turn [Ulrich *et al.* (1995), *supra*] is conserved among bacterial superantigens and may
25 provide the key determinant (hydrophobic or otherwise) for binding HLA-DR. However, TSST-1 does not have a highly charged residue in the polar pocket that interacts with Lys39 of the HLA DRα chain and uses an alternative conformational binding mode that allows
30 TSST-1 to interact with HLA DR1 β-chain residues and the carboxy-terminal region of the antigenic peptide.

Both SEA and SEE bind to the β subunit of DR by means of a single zinc atom [*Fraser, J.D. et al.* (1992) *Proc. Natl. Acad. Sci. USA* **89**, 5507-5511]. The
35 amino-terminal domain of SEA interfaces with the HLA

DRα chain [Ulrich, *et al*. (1995)], while SEA C-terminal domain residues His187, His225 and Asp227 form a zinc-coordination complex, likely with His-81 from the β chain of an adjoining HLA DR molecule.

5   However, our results have shown that binding of superantigen to the HLA DRβ subunit does not directly stimulate T cells [Ulrich *et al*. (1995) *Nature, Struct. Biol.* 2, 554-560], but increases the potential of the bound SEA to interact with the α chain of

10   another HLA DR, thus increasing the biological potency.

     A least-squares superimposition of the unbound molecules of modeled SEA and the crystal structure of SEB, aligned according to their structurally conserved

15   α-helical and ß-strand regions, exhibited a global folding pattern which is very similar. Differences between the two structures are calculated to be located primarily in loops of low sequence homologies, with the largest positional deviations occurring

20   between structurally conserved regions of residues 18-20, 30-32, 173-181, 191-194, and the cysteine-loop region (90-111). Only one of these regions in SEB makes significant contact (residue Y94 [Y=tyrosine] in particular) with the HLA-DR1 molecule [Jardetzky, T.S.

25   *et al*. (1994) *Nature* **368**, 711-718].

     The binding interface between SEB and HLA-DR1 consists principally of two structurally conserved surfaces located in the N-terminal domain: a polar binding pocket derived from three ß-strand elements of

30   the ß-barrel domain and a hydrophobic reverse turn. The binding pocket of SEB contains residues E67 (E=Glutamic acid), Y89 (Y=Tyrosine) and Y115 (Y=tyrosine), and binds K39 (K=Lysine) of the DRα subunit. The amino acid one letter code is defined as

35   the following: A= Alanine (Ala), I= Isoleucine (Ile),

L= Leucine (Leu), M= Methionine (Met), F=
Phenylalanine (Phe), P= Proline (Pro), W=Tryptophan
(Trp), V=Valine (Val), N= Asparagine (Asn), C=Cysteine
(Cys), Q= Glutamine (Q), G= Glycine (Gly), S= Serine

5   (Ser), T= Threonine (Thr), Y= Tyrosine (Tyr), R=
Arginine (Arg), H=Histidine (His), K= Lysine (Lys), D=
Aspartic acid (Asp), and E= Glutamic acid (Glu). For
SEA, the binding interface with the DR molecule is
modeled to contain a similar binding pocket consisting

10   of residues D70, Y92 and Y108. Mutation of residue
Y89 in SEB or Y92 in SEA to alanine (**Fig. 2**) resulted
in greater than 100-fold reduction in DR1 binding.
The substitution of alanine for Y89 in SEB and Y92 in
SEA eliminates the hydrogen bond with K39 and disrupts

15   packing interactions with adjacent protein residues.
Modeling of the SEA mutant Y92A predicts an increase
in solvent-accessible surface area for Y108 by a
factor of two greater than the wild-type structure,
allowing the formation of a hydrogen bond to the

20   carboxylate group of D70 and thus disrupting key
anchoring and recognition points for HLA-DR1. This
effect is expected to be somewhat less in SEB due to
the longer side chain at E67. Substitution of SEB
Y115 with alanine also resulted in greater than 100-

25   fold reduction of binding. In contrast, the same
replacement of Y108 in SEA yielded little to no change
in DR1 binding (**Fig. 2a**), suggesting the primary
importance of SEA residues Y92 and D70 for stabilizing
interactions with K39. The K39 side chain of DRα

30   forms a strong ion-pair interaction with the SEB E67
carboxylate group and hydrogen bonds with the hydroxyl
groups of Y89 and Y115. Substitution of SEB E67 by
glutamine reduced binding affinity by greater than
100-fold (**Fig. 2**), reflecting the replacement of the

35   strong ionic bond with a weaker hydrogen bond. To

optimize ion-pair interactions of the analogous SEA
site, the shorter carboxylate side chain of D70 is
predicted to shift K39 of DRα, weakening interactions
with SEA Y108. The substitution of alanine for SEA

5   Y108 is thus more easily accommodated than the
homologous substitution of SEB Y115, without loss in
DR1 binding.

Comparisons of the polar pocket with other
bacterial superantigens were then made. SEC1-3 and

10 SPE-A have conserved the critical DR1 binding-
interface residues (**Fig. 1**), and share with SEB and
SEA secondary structural elements of the DR1-binding
surfaces. Asparagine in SED (N70) replaces the acidic
side chain present in SEA, SEB, SPE-A and SEC1-3.

15 Accordingly, for SED the salt bridge of the polar
pocket is likely to be replaced by a hydrogen bond.
Overall, DR1 affinities for SED and SEA appeared to be
equivalent (**Fig. 2b**), indicating that other
interactions may compensate for the absence in SED of

20 the ion-pair found in the other superantigens. For
the case of TSST-1, mutating DRα residues K39 to
serine or M36 to isoleucine has been shown to greatly
reduce binding [Panina-Bordignon *et al.* (1992) *J. Exp.
Med.* **176**: 1779-1784]. Although primarily

25 hydrophobic, the critical TSST-1 structural elements
are conserved with the SEA and SEB polar binding
pocket. SEB residues Y89 and Y115 are homologous to
T69 and I85 in TSST-1, respectively, and SEB E67 is
replaced by I46. These TSST-1 residues are positioned

30 in a conserved ß-barrel domain found in both SEB and
SEA. However, the TSST-1 site lacks polarity
equivalent to SEB/SEA, and hydrogen bonding with the
hydroxyl of TSST-1 residue T69 would require that DRα
K39 extend 5 Å into the pocket. TSST-1 binding

35 utilizes an alternative strategy [Kim *et al.* (1994)

Science **266**:1870-1874] consisting of hydrophobic contacts centered around residue I46, and potential ionic or hydrogen bonds bridging DRα residues E71 and K67 to R34 and D27, respectively, of TSST-1.

5 The hydrophobic region of the binding interface between SEB and the HLA-DR1 molecule consists of SEB residues 44-47, located in a large reverse turn connecting ß-strands 1 and 2 of SEB. These residues appear to make strong electrostatic interactions with

10 DRα through their backbone atoms. The mutation of L45 to an arginine reduced overall HLA-DR1 binding greater than 100-fold (**Fig. 2b**), attributable to the less energetically favorable insertion of a highly charged residue into a hydrophobic depression on the DR1

15 molecule. The modeled DR1-SEA complex presents similar interactions with the SEA backbone atoms, with the exception of a glutamine (Q49) replacing SEB Y46. Mutation of L48 to glycine in SEA (homologous to L45 of SEB) has been reported to decrease T-cell

20 responses. SEB L45 and the comparable L30 of TSST-1 are the most extensively buried residues in the DR1 interface. The leucine is conserved among the bacterial superantigens (**Fig. 3**) and may provide the necessary hydrophobic structural element for surface

25 complementarity with DR1, consistent with the mutagenesis data for SEB and SEA.

The inventor has performed similar structure and function studies with TSST-1, SEC1 and SPE-A.

In determining the overall affinity of the

30 superantigen for DR1, a contributory role is played by structural variations around the common binding motifs. A short, variable structured, disulfide-bonded loop is found in SEA and a homologous longer loop in SEB. The SEB residue Y94, contained within

35 this loop, forms hydrophobic interactions with L60 and

A61 of the DRα subunit. Replacement of Y94 with
alanine partially inhibits DR1 binding (**Fig. 2a,b**).
An alanine is found in SEA (A97) and SEE at the
position equivalent to SEB Y94, and mutating this
5   residue in SEA to tyrosine results in disrupted
instead of stabilized interactions with DR1 (**Fig.
2a**). Although the disulfide loops differ in structure
between SEA and SEB, A97 apparently contributes to the
DRα binding interface in a manner similar to Y94 of
10   SEB. Because TSST-1 lacks a disulfide loop, similar
contacts with DRα are replaced by interactions with β-
strands of TSST-1. In a like manner, the absence of a
salt bridge between the residues K39 of DRα and N65 of
SED is apparently compensated for by stabilizing
15   interactions occurring outside of the otherwise
conserved dominant binding surfaces (**Fig. 2a**).

The amino acid residues in contact with TCR are
located in regions of high sequence variability,
presenting a unique surface for interaction with the
20   TCR. Residues implicated in TCR interactions by
mutagenesis of SEA and SEB reside in variable loop
regions, while TSST-1 mutants that affect TCR binding
are mainly located in an α helix [Acharya, R.K. *et al*.
(1994) *Nature* **367**, 94-97; Kim, J. *et al*. (1994)
25   *Science* **266**, 1870-1874]. Specifically, mutations that
diminish T-cell receptor recognition of SEB include
residues N23, Y61, and the homologous SEA N25 or Y64
(**Fig. 2c**). SEA residues S206 and N207 also control
T-cell responses [Hudson, *et al*.(1992) *J. Exp. Med.*
30   **177**: 175-184]. Mutants of the polar binding pocket,
SEA Y92A and SEB Y89A, equivalently reduced T-cell
responses (**Fig. 2c**), reflecting the observed
decreases in DR1-binding (**Fig. 2a, b**). While
supporting reduced T-cell responses, mutants SEA Y64A

and SEB Y61A retained normal affinities for DR1 (**Fig. 2a-c**).

In view of the detailed description of the
present invention and the results of molecular
5   modelling and structural studies of staphylococcal and
streptococcal superantigen toxins discussed above, any
amino acid sequence derived from a superantigen toxin
can be altered.   Sequences of several superantigen
toxins are already known and available to the public
10   in sequence databases such as GenBank, for example.
The superantigen toxin sequence is preferably altered
at the hydrophobic loop or polar binding pocket
depending on the superantigen.   Alternatively,
residues adjacent to the hydrophobic loop or polar
15   binding pocket that contact HLA-DR or residues at
sites that can indirectly alter the structure of the
hydrophobic loop or polar pocket can be altered.   The
number of residues which can be altered can vary,
preferably the number can be 1-2, more preferably 2-3,
20   and most preferably 3-4, or more with the limitation
being the ability to analyze by computational methods
the consequences of introducing such mutations.   The
residues which can be altered can be within 5 amino
acid residues of the central Leucine of the
25   hydrophobic loop (such as L45 of SEB), or within 5
residues of one of the amino acid residues of the
polar binding pocket that can contact HLA-DR, (such as
E67, Y89, or Y115 of SEB), more preferably, within 3
amino acid residues of the central Leucine of the
30   hydrophobic loop (such as L45 of SEB), or within 3
residues of one of the amino acid residues of the
polar pocket that can contact HLA-DR, (such as E67,
Y89, or Y115 of SEB), and most preferably, the central
Leucine of the hydrophobic loop (such as L45 of SEB),
35   or one of the amino acid residues of the polar binding

pocket that can contact HLA-DR, (such as E67, Y89, or
Y115 of SEB). The residues can be changed or
substituted to alanine for minimal disruption of
protein structure, more preferably to a residue of
5 opposite chemical characterisitcs, such as hydrophobic
to hydrophilic, acidic to neutral amide, most
preferably by introduction of a residue with a large
hydrated side chain such as Arginine or Lysine. In
addition, side chains of certain nonconserved
10 receptor-binding surfaces, can also be altered when
designing superantigen toxins with low binding
affinities. These residues can include Y94 of SEB and
structurally equivalent residues of other
superantigens, such as A97 of SEA, or any side chain
15 within 5 residues from these positions or any side
chain in discontinuous positions (discontinuous
positions are defined as amino acid residues that fold
together to form part of a discrete three-dimensional
structural unit but are not present on the same
20 secondary structural unit e.g. α helix or β-strand)
such as disulfide-bonded side chains, that involve,
directly or indirectly, the nonconserved receptor
contact surfaces outside of the polar binding pocket
or hydrophobic loop. Further, amino acid residues
25 involved with protein folding or packing can be
altered when designing superantigen toxins with low
binding affinities [Sundstrom *et al.* (1996) *EMBO J.*
15, 6832-6840; Sundstrom *et al.* (1996) *J. Biol. Chem.*
271, 32212-32216; Acharya *et al.* (1994) *Nature* **367**,
30 94-97; Prasad *et al.* (1993) *Biochem.* **32**, 13761-13766;
Swaminathan *et al.* (1992) *Nature* **359**, 801-806].
Furthermore, especially for superantigens with higher
affinities for T-cell antigen receptors, side chains
of amino acids within 5 residues of the position
35 represented by N23 (conserved residue in most

superantigens) , N60 (conserved Asn or Trp in most
superantigens) Y91 (semiconserved hydrophobic residues
Trp, Ile, Val, His in most superantigens) and D210 of
SEB (conserved Asp in most superantigens) can be

5 altered when designing superantigen toxins with low
binding affinities. These residues are likely to form
part of the integral molecular surfaces that are in
contact with T-cell antigen receptors. Because the T-
cell receptor contact areas of superantigen toxins are

10 essential for causing specific activation or
inactivation of T-cell subsets, altering residues that
are unique to each superantigen but that are located
within 5 residues of the positions represented by N23,
N60 and Y91 can produce superantigens that affect a

15 smaller number (e.g. 1-3) of subsets. Such altered
superantigen toxins can be useful as therapeutic
agents.

In another embodiment, the present invention
relates to a DNA or cDNA segment which encodes a

20 superantigen toxin such as SEA, SEB, SEC-1, SPEa, and
TSST-1 to name a few, the sequence of which has been
altered as described above to produce a toxin protein
with altered binding ability to MHC Class II and/or T-
cell receptors. For SEA, the following three

25 mutations were introduced into the toxin molecule:
Tyrosine at amino acid position 92 changed to alanine;
Aspartic acid at amino acid position 70 changed to
arginine; Leucine at amino acid position 48 changed to
arginine. The reduction in binding to HLA DR is

30 additive per mutation, though one or two mutations can
produce a vaccine and a combination of all three
mutations in one molecule produces a better vaccine.
Other substitutions can also result in reduced
binding.

The B899445 vaccine consists of the following three mutations simultaneously introduced into the toxin molecule: tyrosine at amino acid position 89 changed to alanine; tyrosine at amino acid position 94 changed to alanine; leucine at amino acid position 45 changed to arginine. The altered superantigen toxins can be expressed either as a full-length propolypeptide or as a polypeptide in which the leader peptide has been deleted. The full-length expressed product (SEA vaccine, A489270P *(SEQ ID NO:2)*; SEB vaccine B899445P *(SEQ ID NO:8)*, B2360210P *(SEQ ID NO:6)*) is secreted into the periplasmic space of *E. coli* host cells, and the leader peptide is recognized and cleaved by a native bacterial enzymatic mechanism. The altered superantigen toxins in which the leader peptide has been deleted (A489270C *(SEQ ID NO:4)*, B899445C *(SEQ ID NO:10)*), the first residue of the mature protein is encoded by the transcriptional start site and codon for methionine (ATG), and the protein is expressed as a nonsecreted product within the host *E. coli* cell. For the TSST-1 vaccine TST30 *(SEQ ID NO:12)*, the leucine at position 30 was changed to arginine. For the SEC1 vaccine, SEC45 *(SEQ ID NO:14)*, the leucine at position 45 was changed to arginine. For the SPE-A vaccine, SPEA42 *(SEQ ID NO:16)*, the leucine at position 42 was changed to arginine.

In another embodiment, the present invention relates to a recombinant DNA molecule that includes a vector and a DNA sequence as described above. The vector can take the form of a plasmid such as any broad host range expression vector for example pUC18/19, pSE380, pHIL, pET21/24 and others known in the art. The DNA sequence is preferably functionally linked to a promoter such that the gene is expressed when present in an expression system and an altered superantigen toxin is produced. The expression system can be an *in vitro* expression system or host cells

such as prokaryotic cells, or *in vivo* such as DNA vaccines.

In a further embodiment, the present invention relates to host cells stably or transiently

5   transformed or transfected with the above-described recombinant DNA constructs. The host can be any eukaryotic or prokaryotic cell including but not limited in *E. coli* DH5α or BL21. The vector containing the altered superantigen toxin gene is

10   expressed in the host cell and the product of the altered toxin gene, whether a secreted mature protein or a cytoplasmic product, can be used as a vaccine or as a reagent in diagnostic assays or detection methods, or for therapeutic purposes. Please see

15   e.g., Maniatis, Fitsch and Sambrook, Molecular Cloning; A Laboratory Manual (1982) or DNA Cloning, Volumes I and II (D. N. Glover ed. 1985) for general cloning methods. The DNA sequence can be present in the vector operably linked to a highly purified IgG

20   molecule, an adjuvant, a carrier, or an agent for aid in purification of altered toxin. The transformed or transfected host cells can be used as a source of DNA sequences described above. When the recombinant molecule takes the form of an expression system, the

25   transformed or transfected cells can be used as a source of the altered toxin described above.

A recombinant or derived altered superantigen toxin is not necessarily translated from a designated nucleic acid sequence; it may be generated in any

30   manner, including for example, chemical synthesis, or expression of a recombinant expression system. In addition the altered toxin can be fused to other proteins or polypeptides for directing transport for example into the periplasm or for secretion from the

cell. This includes fusion of the recombinant or derived altered superantigen to other vaccines or sequences designed to aid in purification, such as His-tagged, epitope-tagged or antibody Fc-fusions.

5   In a further embodiment, the present invention relates to a method of producing altered superantigen toxin which includes culturing the above-described host cells, under conditions such that the DNA fragment is expressed and a superantigen toxin protein

10   is produced. The superantigen toxin can then be isolated and purified using methodology well known in the art such as immunoaffinity chromatography or preparative isoelectric focusing. However, the method of purification is not critical to the performance of

15   the vaccine. The altered superantigen toxin can be used as a vaccine for immunity against infection with bacterial superantigen toxins or as a diagnostic tool for detection of superantigen toxin-associated disease or bacterial infection. The transformed host cells

20   can be used to analyze the effectiveness of drugs and agents which affect the binding of superantigens to MHC class II or T-cell antigen receptors. Chemically derived agents, host proteins or other proteins which result in the down-regulation or alteration of

25   expression of superantigen toxins or affect the binding affinity of superantigen toxins to their receptors can be detected and analyzed. A method for testing the effectiveness of a drug or agent capable of altering the binding of superantigen toxins to

30   their receptors can be for example computer-aided rational design or combinatorial library screening, such as phage display technology.

     In another embodiment, the present invention relates to antibodies specific for the above-described

35   altered superantigen toxins. For instance, an

antibody can be raised against the complete toxin or against a portion thereof. Persons with ordinary skill in the art using standard methodology can raise monoclonal and polyclonal antibodies to the altered

5 superantigens of the present invention, or a unique portion of the altered superantigen. Materials and methods for producing antibodies are well known in the art (see for example Goding, in, Monoclonal Antibodies: Principles and Practice, Chapter 4, 1986).

10 The antibodies can be used in diagnostic assays for detection of superantigen toxin-associated infection. Neutralizing antibodies can be used in a therapeutic composition for the treatment of amelioration of anergy and/or for the treatment of a superantigen

15 toxin-associated infection.

In a further embodiment, the present invention relates to a method for detecting the presence of superantigen-associated bacterial infections in a sample. Using standard methodology well known in the

20 art, a diagnostic assay can be constructed by coating on a surface (i.e. a solid support) for example, a microtitration plate or a membrane (e.g. nitrocellulose membrane), all or a unique portion of the altered superantigen described above, and

25 contacting it with the serum of a person suspected of having a superantigen-associated bacterial infection. The presence of a resulting complex formed between the altered superantigen toxin and antibodies specific therefor in the serum can be detected by any of the

30 known methods common in the art, such as fluorescent antibody spectroscopy or colorimetry. This method of detection can be used, for example, for the diagnosis of superantigen-associated bacterial infections.

In yet another embodiment, the present invention relates to a method for detecting the presence of superantigen toxin in a sample. Using standard methodology well known in the art, a diagnostic assay

5 can be constructed by coating on a surface (i.e. a solid support) for example, a microtitration plate or a membrane (e.g. nitrocellulose membrane), antibodies specific for altered superantigen toxin, and contacting it with serum or tissue sample of a person

10 suspected of having superantigen-associated bacterial infection. The presence of a resulting complex formed between toxin in the serum and antibodies specific therefor can be detected by any of the known methods common in the art, such as fluorescent antibody

15 spectroscopy or colorimetry. This method of detection can be used, for example, for the diagnosis of superantigen-associated bacterial infection or disease such as food poisoning and toxic-shock syndrome or the detection of superantigen toxin in food and drink.

20 In another embodiment, the present invention relates to a diagnostic kit which contains altered superantigen toxin from a specific bacteria or several different superantigen toxins from bacteria and ancillary reagents that are well known in the art and

25 that are suitable for use in detecting the presence of antibodies to superantigen toxin-associated bacteria in serum or a tissue sample. Tissue samples contemplated can be avian, fish, or mammal including monkey and human.

30 In yet another embodiment, the present invention relates to a vaccine for protection against superantigen toxin-associated bacterial infections. The vaccine can comprise one or a mixture of individual altered superantigen toxins, or a portion

35 thereof. When a mixture of two or more different

altered superantigen toxin from different bacteria is used, the vaccine is referred to as a multivalent bacterial superantigen vaccine. The vaccine is designed to protect against the pathologies resulting

5 from exposure to one or several related staphylococcal and streptococcal toxins. In addition, the protein or polypeptide can be fused or absorbed to other proteins or polypeptides which increase its antigenicity, thereby producing higher titers of neutralizing

10 antibody when used as a vaccine. Examples of such proteins or polypeptides include any adjuvants or carriers safe for human use, such as aluminum hydroxide.

The staphylococcal enterotoxin (SE) serotypes

15 SEA, SED, and SEE are closely related by amino acid sequence, while SEB, SEC1, SEC2, SEC3, and the streptococcal pyrogenic exotoxins B share key amino acid residues with the other toxins, but exhibit only weak sequence homology overall. However, there are

20 considerable similarities in the known three-dimensional structures of SEA, SEB, SEC1, SEC3, and TSST-1. Because of this structural similarity, it is likely that polyclonal antibodies obtained from mice immunized with each SE or TSST-1 exhibit a low to high

25 degree of cross-reaction. In the mouse, these antibody cross-reactions are sufficient to neutralize the toxicity of most other SE/TSST-1, depending upon the challenge dose. For example, immunization with a mixture of SEA, SEB, TSST-1 and SPEa was sufficient to

30 provide antibody protection from a challenge with any of the component toxins, singly or in combination.

The likelihood of substantial antigen-cross-reactivity suggests that it may be possible to obtain immune protection for other (or perhaps all)

35 staphylococcal superantigens by use of a minimal mixed

composition of vaccines. For the case of
staphylococcal superantigens, a combination of the
component vaccines from SEA, SEB, SEC-1 and TSST-1
should be sufficient to provide immune protection

5   against SEA, SEB, SEC1-3, and TSST-1. The addition of
SPEa component to the trivalent mixture will allow for
sufficient protection against the streptococcal toxins
SPEa and SPEc. Therefore, a multivalent vaccine
consisting of the altered superantigen toxins from

10 SEA, SEB, SEC-1, TSST-1, and SPEa as described above,
is predicted to provide protective immunity against
the majority of bacterial superantigen toxins.
        The vaccine can be prepared by inducing
expression of a recombinant expression vector

15 comprising the gene for the altered toxin described
above. The purified solution is prepared for
administration to mammals by methods known in the art,
which can include filtering to sterilize the solution,
diluting the solution, adding an adjuvant and

20 stabilizing the solution. The vaccine can be
lyophilized to produce a vaccine against superantigen
toxin-associated bacteria in a dried form for ease in
transportation and storage. Further, the vaccine may
be prepared in the form of a mixed vaccine which

25 contains the altered superantigen toxin(s) described
above and at least one other antigen as long as the
added antigen does not interfere with the
effectiveness of the vaccine and the side effects and
adverse reactions, if any, are not increased

30 additively or synergistically. Furthermore, the
vaccine may be administered by a bacterial delivery
system and displayed by a recombinant host cell such
as *Salmonella* spp, *Shigella* spp, *Streptococcus* spp.
Methods for introducing recombinant vectors into host

35 cells and introducing host cells as a DNA delivery

system are known in the art [Harokopakis *et al.* (1997) *Infect. Immun.* **65**, 1445-1454; Anderson *et al.* (1996) *Vaccine* **14**, 1384-1390; Medaglini *et al.* (1995) *Proc. Natl. Acad. Sci. U.S.A.* **92**, 6868-6872].

5   The vaccine may be stored in a sealed vial, ampule or the like. The present vaccine can generally be administered in the form of a liquid or suspension. In the case where the vaccine is in a dried form, the vaccine is dissolved or suspended in sterilized

10 distilled water before administration. Generally, the vaccine may be administered orally, subcutaneously, intradermally or intramuscularly but preferably intranasally in a dose effective for the production of neutralizing antibody and protection from infection or

15 disease.

In another embodiment, the present invention relates to a method of reducing superantigen-associated bacterial infection symptoms in a patient by administering to said patient an effective amount

20 of anti-altered superantigen toxin antibodies, as described above. When providing a patient with anti-superantigen toxin antibodies, or agents capable of inhibiting superantigen function to a recipient patient, the dosage of administered agent will vary

25 depending upon such factors as the patient's age, weight, height, sex, general medical condition, previous medical history, etc. In general, it is desirable to provide the recipient with a dosage of the above compounds which is in the range of from

30 about 1pg/kg to 10 mg/kg (body weight of patient), although a lower or higher dosage may be administered.

In a further embodiment, the present invention relates to a therapeutic method for the treatment of diseases that may not be associated directly with

35 superantigen toxins but which result in specific

nonresponsiveness of T-cell subsets or detection of abnormally low level of subsets in peripheral blood, said method comprising the administration of altered superantigen toxins, *in vivo* or *ex vivo*, such that T-

5   cell subsets are expanded or stimulated. Diseases which cause anergy or nonresponsiveness of T-cells include, but are not limited to, infectious diseases and cancers. The desired clinical outcome such as an increase in detectable T cell subsets or in

10   stimulation *ex vivo* of T-cells through their antigen receptors, such as by antigen or anti-CD3 antibody can be measured by standard clinical immunology laboratory assays.

In yet another embodiment, the present invention

15   relates to a therapeutic method for the treatment of diseases associated with expanded or over-stimulated T-cell subsets, such as autoimmunity for example, said method comprising administration *in vivo* or *ex vivo*, of superantigen toxin altered in such a manner that

20   only limited (1-3) T-cell subsets are stimulated but that MHC class II binding affinity still remains, such that anergy or inactivation of T-cells is produced. The desired clinical outcome can be measured as a reduction of circulating blood T-cells of the targeted

25   subset(s) or diminished antigen or other antigen receptor-mediated-stimulatory responses by assays known in the art.

Described below are examples of the present

30   invention which are provided only for illustrative purposes, and not to limit the scope of the present invention.   In light of the present disclosure, numerous embodiments within the scopy of the claims will be apparent to those of ordinary skill in the

35   art.

The following Materials and Methods were used in the Examples that follow.

Structural comparisons

Primary protein structure data are available for
5   several bacterial superantigens, including SEA, SED, SEB, SEC1-3, TSST-1. Superantigens for which structures were unavailable were modeled using comparative techniques (HOMOLOGY program; Biosym Technologies, Inc., San Diego, CA). Before x-ray
10   crystallography data was available, SEA was modeled by using this method, and the model was in very close agreement with the experimentally determined structure. As an example, the amino acid sequence for SEA was aligned with the known structure of free and
15   HLA-DR1 bound SEB, and the SEA molecule was built for both free and DR1-bound proteins. Loop segments of SEA were generated by a de novo method. Refinement of the modeled structures was carried out by means of molecular-dynamics simulations (DISCOVER, Biosym).
20   The constructed free SEA molecule was immersed in a 5-Å layer of solvent water and the α-carbon atoms lying in the structurally conserved regions were tethered to their initial positions during the simulations. For the bound SEA molecule, simulations were carried out
25   by constructing an active-site region composed of part of the SEA molecule and the DR1 molecule inside a 10-Å interface boundary, as derived from the crystal structure of the DR1-SEB complex. Amino acid residues lying in the outer boundary were rigidly restrained at
30   their initial positions. The active-site region was immersed in a 5-Å layer of water. Protein interactions were modeled by employing the consistent valence force field with a non-bonded cutoff distance of 11.0 Å. Simulations were initiated with 100 cycles
35   of minimization using a steepest descent algorithm

followed by 100-ps relaxation (using a 1.0 fs timestep). Structural comparisons between SEB, SEC1, and TSST-1 were performed by using the crystal structures (Brookhaven data holdings) aligned

5   according to common secondary structural elements and/or by sequence and structural homology modeling.

Site-specific mutagenesis

Site-specific mutagenesis was performed according

10   to the method developed by Kunkel, using gene templates isolated from Staphylococcus aureus strains expressing SEA (FDA196E, a clinical isolate, Fraser, J.D. (1994) *Nature* **368**: 711-718), SEB (14458, clinical isolate), SEC1 (Toxin Technologies, Sarasota,

15   FL), TSST-1 (pRN6550 cloned product, a clinical isolate, Kreiswirth, B. N. *et al.* (1987) *Mol. Gen. Genet.* **208**, 84-87), and SPEa (Toxin Technologies), respectively. Modified T7 polymerase (Sequenase, U.S. Biochemical Corp., Cleveland, OH) was used to

20   synthesize second-strand DNA from synthetic oligonucleotides harboring the altered codon and single-stranded, uracil-enriched M13 templates. Mutagenized DNA was selected by transforming *E. coli* strain JM101. Alternatively, double stranded DNA was

25   used as template for mutagenesis. Mutagenized sequences were confirmed by DNA sequencing (Sanger *et al.*, 1977, *Proc. Natl. Acad. Sci. USA* **74**: 5463-5467; Sambrook *et al.*, 1989) using synthetic primers derived from known sequences, or universal primers. The

30   complete coding sequences were inserted into expression plasmids such as pUC19, pSE380 or pET21 for production in *E. coli* hosts.

Protein purifications

The appropriate *E. coli* hosts were transformed with plasmids harboring the mutant toxin genes. In general, the bacteria were grown to an A600 0.5-0.6 in

5   Terrific Broth (Difco Laboratories, Detroit, MI) containing 50 µg/mL ampicillin or kanamycin. Recombinant proteins were induced with isopropyl-ß-D-thio-galactopyranoside (Life Technologies, Gaithersburg, MD) and recovered as cytoplasmic or

10 bacterial periplasmic secretion products. Bacteria were collected by centrifugation, washed with 30 mM NaCl, 10 mM TRIS (pH 7.6), and pelleted by centrifugation and either lysed or osmotically shocked for collection of secreted proteins. Preparations

15 were isolated by CM Sepharose ion-exchange chromatography, rabbit antibody (Toxin Technologies, Sarasota, FL) affinity columns, ion exchange HPLC or similar methods. In some cases partially purified superantigen was further purified by preparative

20 isoelectric focusing (MinipHor; Rainin Instrument Company, Inc., Woburn, MA.). The MinipHor was loaded with the SEA-enriched fraction from CM Sepharose chromatography in a solution containing 10% (v/v) glycerol and 1% (v/v) pH 6-8 ampholytes (Protein

25 Technologies, Inc., Tucson, AZ). The protein preparations were allowed to focus until equilibrium was reached (approximately 4 hr, 4°C). Twenty focused fractions were collected and aliquots of each were analyzed by SDS-polyacrylamide gel electrophoresis

30 (SDS-PAGE) and immunoblotting. The SEA-containing fractions were pooled, and refocused for an additional 4 h. The fractions containing purified SEA were pooled and dialyzed first against 1 M NaCl (48 h, 4°C) to remove ampholytes, and then against PBS (12 h,

35 4°C). Legitimate amino-terminal residues were

confirmed by protein sequencing. Precise measurements of protein concentrations were performed by immunoassay using rabbit antibody affinity-purified with the wild-type superantigens and by the

5   bicinchoninic acid method (Pierce, Rockford, IL) using wild-type protein as standards. All protein preparations were >99% pure, as judged by SDS-PAGE and Western immunoblots. In some cases, as when used for lymphocyte assays, bacterial pyrogens were removed by

10   passing the protein preparations over Polymyxin B affinity columns.

Binding of superantigens to HLA-DR1

The DR1 homozygous, human B-lymphoblastoid cell

15   line LG2 or L cells transfected with plasmids encoding HLA-DR1αβ were used in the binding experiments. Cells were incubated 40 min (37°C) with wild-type or mutant superantigen in Hanks balanced salt solution (HBSS) containing 0.5% bovine serum albumin. The cells were

20   washed with HBSS and then incubated with 5 μg of specific rabbit antibody (Toxin Technology, Sarasota, FL) for 1 h on ice. Unbound antibody was removed, and the cells were incubated with FITC-labelled goat anti-rabbit IgG (Organon Teknika Corp., Durham, N.C.) on

25   ice for 30 min. The cells were washed and analyzed by flow cytometry (FACScan; Becton Dikinson & Co., Mountain View, CA). Controls consisted of cells incubated with affinity purified anti-toxin and the FITC labelled antibody without prior addition of

30   superantigen.

Lymphocyte proliferation

Human peripheral blood mononuclear cells were purified by Ficoll-hypaque (Sigma, St. Louis, MO)

buoyant density gradient centrifugation. Genes encoding the human MHC class II molecules DR1$\alpha\beta$ (DRA and DRB1*0101 cDNA [Bavari and Ulrich (1995) *Infect. Immun.* **63**, 423-429] were cloned into the eukaryotic

5 expression vector pRC/RSV (Invitrogen, Carlsbad, CA), and mouse L cells were stably transfected. The transfectants were selected by fluorescence-activated cell sorting (EPICS C, Coulter Corp., Hialeah, FL) using rabbit anti-DR$\alpha\beta$ antisera and FITC-goat anti-

10 rabbit IgG, to produce cells that expressed a high level of DR$\alpha\beta$21. 1×10^5 cells/well of a 96-well plate were irradiated (15,000 Rad), and wild-type or mutant SE, was added. After a brief incubation period (45 min, 37°C), unbound SE was rinsed from the culture

15 plates using warm media. The cells were cultured in RPMI-1640 (USAMRIID) with 5% FBS for 72 h, and pulsed-labelled for 12 h with 1μCi [^{3}H]-thymidine (Amersham, Arlington Heights, IL). Cells were harvested onto glass fiber filters, and [^{3}H]-thymidine incorporation

20 into the cellular DNA was measured by a liquid scintillation counter (BetaPlate, Wallac Inc., Gaithersburg, MD). Splenic mononuclear cells or human peripheral blood mononuclear cells were obtained by buoyant density centrifugation (Histopaque; Sigma

25 Chemical Comp.) and washed three times. The cells were resuspended in medium containing 5% fetal bovine serum (FBS), and 100 μl (4×10^5 cells) of the cell suspension was added to triplicate wells of 96-well flat bottom plates. The mononuclear cells were

30 cultured (37°C, 5% CO_2) with WT or mutant SEA. After 3 days the cultures were pulsed (12h) with 1 μCi/well of [^{3}H]thymidine (Amersham, Arlington Heights, IL) and incorporated radioactivity was measured by liquid scintillation.

Gel electrophoresis and immunoblotting analysis.

The protein preparations were analyzed by SDS-PAGE (12%) and stained with Coomassie Brilliant Blue

5   R-250 (Sigma Chemical Comp. St Louis, MO) in methanol (10% v/v) acetic acid (10% v/v). The proteins separated by SDS-PAGE (not stained) were transferred to nitrocellulose membranes (Bio-Rad Lab. Inc., Melville, NY) by electroblotting, and the membranes

10 were then blocked (12 h, 4°C) with 0.2% casein in a buffer consisting of 50 mM sodium phosphate, 140 mM sodium chloride, pH 7.4 (PBS). The membrane was then incubated (1 h, 37°C, shaking) with 2 µg/mL of affinity-purified anti-toxin antibody (Toxin

15 Technology, Sarasota, FL) in PBS with 0.02% casein. After the membranes were thoroughly washed, peroxidase-conjugated goat anti-rabbit IgG (Cappel/Organon Teknika Corp., West Chester, PA) was added (1:5,000) and the membranes were incubated for 1

20 h (37°C) with shaking. The unbound antibody was removed by washing with PBS and bound antibody was visualized by using a Bio-Rad peroxidase development kit (Biorad, Hercules, CA). For quantitation, dilutions of wild-type preparations were immobilized

25 on nitrocellulose membranes by using a Slot-Blot apparatus (Bio-Rad). The membrane was removed from the Slot-Blot apparatus and unreacted sites were blocked (12h, 4°C) with 0.2% casein in PBS. After washing once with the PBS, the membrane was incubated

30 (1h, 37°C) with 2 µg/mL rabbit affinity purified anti-toxin antibody (Toxin Technology) in PBS that contained 0.02% casein. After four washes, the bound rabbit antibody was reacted with goat anti-rabbit IgG conjugated with horseradish peroxidase (1 h, 37°C) and

35 the blots were developed using enhanced

chemiluminescence (ECL; Amersham Life Sciences,
Arlington Heights, IL) or similar methods. The amount
of mutant protein was measured by densitometry (NIH
Image 1.57 software, National Institutes of Health,

5   Bethesda, MD) of exposed X-ray film. Standard curves
were prepared by plotting the mean of duplicate
densitometric readings for each dilution of toxin
standard. The resulting values were fitted to a
straight line by linear regression. Concentrations of

10   proteins were determined by comparing mean values of
various dilutions of the mutant to the standard curve.

Biological activities and Immunizations.

Male C57BL/6 mice, 10 to 12-weeks old, were

15   obtained from Harlan Sprague-Dawley, Inc. (Frederick
Cancer Research and Development Center, Frederick,
MD). The lethal effect of WT or mutant SEA was
evaluated as described in Stiles *et al.* (1993) *Infect.
Immun.* **61**, 5333-5338. For immunizations, mice were

20   given by interperitoneal (ip) injections either 2 or
10 µg of WT or mutant toxin in 100 µl of adjuvant
(RIBI, Immunochem Research, Inc. Hamilton, MT or
alum), or adjuvant only, and boosted (ip) at 2 and 4
weeks. Serum was collected from tail veins one week

25   after the last immunization. Mice were challenged 2
weeks after the last injection with toxin and
lipopolysaccharide (LPS, 150 µg) from *E. coli* 055:B5
serotype (Difco Laboratories, Detroit, MI). Challenge
controls were adjuvant-immunized or non-immunized mice

30   injected with both agents (100% lethality) or with
either wild type toxin or LPS. No lethality was
produced by these negative controls. Monkeys were
immunized with the antigen in the right leg, caudal
thigh muscles. Each received three intramuscular

35   immunizations with a superantigen vaccine plus

adjuvant. Control monkeys received 0.5 ml total
volume of adjuvant (Alhydrogel, Michigan Department of
Public Health) and sterile PBS using the same
techniques and equipment as the immunized monkeys.
5   Immunizations were administered 28±2 days apart and
consisted of 20 µg of the vaccine in adjuvant in a
total volume of 0.5 ml. Immunizations were
administered on day 0, 28±2, and 56±2 using a 23-27 ga
1/2-5/8" needle attached to a 1 ml tuberculin syringe
10 into the caudal thigh.

Antibody assay.

Microtiter plates were coated with 1 µg/well of
WT toxin in 100 µl of PBS (37°C, 2 h). After antigen
15 coating, the wells were blocked with 250 µl of casein
0.2% in PBS for 4 h at 37°C and then washed four times
with PBS containing 0.2% Tween 20. Immune or
nonimmune sera were diluted in PBS containing 0.02%
casein and 100 µl of each dilution was added to
20 duplicate wells. After each well was washed four
times, bound antibody was detected with horse radish
peroxidase (Sigma Chemical Comp., St. Louis, MO)
labelled goat anti-species specific IgG (37°C, 1 h),
using O-phenylenediamine as the chromogen. Mean of
25 duplicates OD (absorbance at 490 nm) of each treatment
group was obtained and these data were compared on the
basis of the inverse of the highest serum dilution
that produced an OD reading four times above the
negative control wells. For negative controls,
30 antigen or serum was omitted from the wells.

Superantigen binding and TCR subset analysis.

Cells from the mouse B-lymphoma line A20 (ATCC,
Rockville, MD)(2-4 x 10^5 cells) were incubated (40 min

at 37°C) with WT or mutant toxin in Hanks balanced salt solution containing 0.5% bovine serum albumin (HBSS, USAMRIID). The cells were washed with HBSS and incubated with 5 µg of affinity-purified anti-toxin

5   antibody in HBSS (4°C, 45 min). Unbound antibody was removed and the bound antibody was detected with fluorescein isothiocyanate (FITC)-labelled, goat anti-rabbit IgG (Organon Teknika Corp., Durham, NC). Unbound antibody was removed and the cells were

10 analyzed by with a FACSort flow cytometer (Becton Dikinson & Co. , Mountain View, CA).

For TCR subset analysis, splenic mononuclear cells were obtained from mice immunized with WT or mutant toxin. The mononuclear cells were incubated

15 (37°C) with WT toxin (100 ng/mL) for 5 days and then cultured in 85% RPMI-1640, 10% interleukin-2 supplement (Advanced Biotechnologies Inc., Columbia, MD) with 5% FBS for an additional 5 days. The T cells were washed twice and stained with anti-TCR

20 (Biosource, Camarillo, CA) or anti-Vß specific TCR (Biosource, Camarillo, CA) (45 min, 4°C). All cells analyzed were positive for T cell marker CD3+ and expressed the CD25 activation marker (data not shown). Controls were incubated with an isotype matched

25 antibody of irrelevant specificity. Unreacted antibody was removed, and the cells were incubated with an FITC-labelled, anti-mouse IgG (Organon Teknika Corp, Durham, NC) on ice for 30 min. The cells were washed and analyzed by flow cytometry (FACSort).

30

LPS potentiation of SE toxicity in mice.

C57BL/6 or BALB/c mice weighing 18-20 g (Harlan Sprague Dawley, Inc., Frederick Cancer Research and Development Center, Frederick, MD) were each injected

35 intraperitoneally (i.p.) with 200 µl of PBS containing

varying amounts of SEA, SEB, or SEC1, TSST-1, or SPEa followed 4 h later with 75 or 150 µg of LPS (200 µl/i.p.). Controls were each injected with either SE (30 mg) or LPS (150 mg). Animals were observed for 72

5   h after the LPS injection. Calculations of LD50 were done by Probit analysis using 95% fiducial limits (SAS Institute Inc., Cary, NC).

The biological effects of SEA and SEB were also tested in transgenic C57BL/6 mice (GenPharm

10   International, Mountain View, CA) deficient in MHC class I or II expression [Stiles *et al.* (1993) *Infect. Immun.* **61**, 5333-5338], as described above, using a single dose of toxin (30 µg/mouse). Genetic homozygosity was confirmed by Southern analysis of

15   parental tail DNA, using β2 microglobulin and MHC class II β DNA probes.

Detection of cytokines in serum.

Mice (n=18 per group) were injected with toxin

20   (10 µg), LPS (150 µg), or toxin plus LPS. Sera were collected and pooled from three mice per group at each time point (2, 4, 6, 8, 10, 22 h) after LPS injection. Sera were collected at various time points following toxin injection (-4 h, or 4h before LPS injection, for

25   data tabulation). Collection of LPS control sera began at the time of injection (0 h).

Serum levels of TNFα and IL-α were detected by an enzyme linked immunosorbent assay (ELISA). TNFα was first captured by a monoclonal antibody against mouse

30   TNFα (GIBCO-BRL, Grand Island, NY) and then incubated with rabbit anti-mouse TNFα antibody (Genzyme, Boston, MA). The ELISA plate was washed and peroxidase conjugate of anti-rabbit antibody (Boehringer Mannheim, Indianapolis, IN) added to the wells. After

35   washing the plate and adding substrate (Kirkegaard and

Perry, Gaithersburg, MD), TNFα concentrations were measured using the mean A450 reading of duplicate samples and a standard curve generated from recombinant mouse TNFα (GIBCO-BRL). Serum levels of

5 IL-1α were determined from the mean reading of duplicate samples with an ELISA kit that specifically detects murine IL-1α (Genzyme, Boston, MA). The standard error of the mean (SEM) for TNFα and IL-1α readings was +/- 5%.

10   Quantitation of IL-6 and IFNγ were measured by bioassays [See et al.(1990) *Infect. Immun.* **58**: 2392-2396]. An IL-6 dependent cell line, 7TD1 (kindly provided by T. Krakauer), was used in a proliferative assay with serial two-fold dilutions of serum samples

15 assayed in triplicate. Proliferation of 7TD1 cells in a microtiter plate was measured by uptake of [^{3}H]-thymidine (1 μCi/well; Amersham, Arlington Heights, IL) and the activity of IL-6 from serum was compared to a recombinant mouse IL-6 standard (R and D Systems,

20 Minneapolis, MN) as previously described [See et al.(1990) *Infect. Immun.* **58**: 2392-2396]. The SEM of triplicate samples was +/- 10%.

     IFNγ was measured by the reduction of vesicular stomatitis virus (New Jersey strain) cytopathic

25 effects on L929 cells, as previously described [Torre et al.(1993) *J. Infect. Dis.* **167**, 762-765]. Briefly, serial two-fold dilutions of serum were made in duplicate and added to microtiter wells containing L929 cells (5 x 10^4/well). After incubating 24 h,

30 virus (5 x 10^5 PFU/well) was added and the cytopathic effects measured at 48 h by absorbance readings (570 nm) of reduced 3-[4, 5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide (Sigma). The activity of each serum sample was determined using recombinant

mouse IFNγ as a standard (Biosource, Camarillo, CA).
The SEM of duplicate samples was +/- 5%.

EXAMPLE 1

5   Molecular modelling and structural studies of
staphylococcal and streptococcal superantigens:
bacterial superantigens share common 3-dimensional
structure.

       Comparison of amino acid sequences (**Fig. 1**)
10   suggested that bacterial superantigens fall into
groups consisting of (1) SEA, SED and SEE, (2) SEB,
staphylococcal enterotoxins C1-C3 (SEC1-3), the
streptococcal pyrogenic exotoxins A (SPE-A) and C
(SPE-C), (3) TSST-1 and (4) the exfoliative toxins
15   (ETA, ETB) and streptococcal pyrogenic exotoxin B
(SPE-B), which are the most distant from the others in
sequence. Although not available to the inventor when
the inventions were first conceived and proof of
principle was obtained, the x-ray crystallographic
20   structures of several bacterial superantigens are now
known. Diverse superantigens, such as SEB and TSST-1,
appear to have little sequence in common, yet they
exhibit homologous protein folds composed largely of β
strands [Prasad, G.S. *et al.* (1993) *Biochemistry* **32**,
25   13761-13766; Acharya, R.K. *et al.* (1994) *Nature* **367**,
94-97; Swaminathan, S. *et al.* (1992) *Nature* **359**, 801-
806]within two distinct domains. Differences between
the proteins are located primarily in highly variable
regions comprised of several surface loops, such as
30   the disulfide-bonded loop which is absent from TSST-1
and at the amino terminus.

       The X-ray crystal structures of SEB and TSST-1
complexed with HLA DR1 are known [Kim, J. *et al.*
(1994) Science 266, 1870-1874 ; Jardetzky, T.S. *et al.*

(1994) Nature 368, 711-718] and this data was useful
to fully explain our results concerning attenuation of
the superantigens by site-specific mutagenesis. The
region of HLA DR1 that contacts SEB consists

5   exclusively of α subunit surfaces. The main regions
of SEB involved are two conserved sites: a polar
pocket derived from three β strands of the β barrel
domain and a highly solvent-exposed hydrophobic
reverse turn. The polar binding pocket of SEB

10 contains a glutamate and two tyrosines that
accommodate Lys39 of the α subunit of HLA DR1, while
the hydrophobic region consists of a leucine and
flanking residues that make several contacts with the
HLA DRα chain. The HLA DR1 binding sites of both

15 TSST-1 and SEB overlap significantly. The hydrophobic
binding contacts of other SAg with the HLA DRα chain
have been proposed [Ulrich et al. (1995) Nature,
Struct. Biol. 2, 554-560] to be similar to those found
in SEB and TSST-1. A motif consisting of a leucine in

20 a reverse turn [Ulrich et al. (1995), supra] is
conserved among bacterial superantigens and may
provide the key determinant (hydrophobic or otherwise)
for binding HLA-DR. However, TSST-1 does not have a
highly charged residue in the polar pocket that

25 interacts with Lys39 of the HLA DRα chain and uses an
alternative conformational binding mode that allows
TSST-1 to interact with HLA DR1 β -chain residues and
the carboxy-terminal region of the antigenic peptide.

Both SEA and SEE bind to the β subunit of DR by

30 means of a single zinc atom [Fraser, J.D. et al.
(1992) Proc. Natl. Acad. Sci. USA 89, 5507-5511]. The
amino-terminal domain of SEA interfaces with the HLA
DRα chain [Ulrich et al. (1995), supra], while SEA C-
terminal domain residues His187, His225 and Asp227

35 form a zinc-coordination complex, likely with His-81

from the β chain of an adjoining HLA DR molecule.
However, our results have shown that binding of
superantigen to the HLA DRβ subunit does not directly
stimulate T cells [Ulrich *et al.* (1995), supra] but

5   increases the potential of the bound SEA to interact
with the α chain of another HLA DR, thus increasing
the biological potency.

EXAMPLE 2

10   Molecular modelling and structural studies of
staphylococcal and streptococcal superantigens: A
detailed protein structure analysis of SEB and SEA
suggested that all bacterial superantigens have a
common mechanism for binding MHC class II receptors.

15   A least-squares superimposition of the unbound
molecules of modeled SEA and the crystal structure of
SEB, aligned according to their structurally conserved
α-helical and ß-strand regions, exhibited a global
folding pattern which is very similar. Differences

20   between the two structures are calculated to be
located primarily in loops of low sequence homologies,
with the largest positional deviations occurring
between structurally conserved regions of residues 18-
20, 30-32, 173-181, 191-194, and the cysteine-loop

25   region (90-111). Only one of these regions in SEB
makes significant contact (residue Y94 in particular)
with the HLA-DR1 molecule [Jardetzky, T.S. *et al.*
(1994) *Nature* **368**, 711-718].

The binding interface between SEB and HLA-DR1

30   consists principally of two structurally conserved
surfaces located in the N-terminal domain: a polar
binding pocket derived from three ß-strand elements of
the ß-barrel domain and a hydrophobic reverse turn.
The binding pocket of SEB contains residues E67, Y89

and Y115, and binds K39 of the DRα subunit. For SEA, the binding interface with the DR molecule is modeled to contain a similar binding pocket consisting of residues D70, Y92 and Y108. Mutation of residue Y89

5   in SEB or Y92 in SEA to alanine (**Fig. 2**) resulted in 100-fold reduction in DR1 binding. The substitution of alanine for Y89 in SEB and Y92 in SEA eliminates the hydrogen bond with K39 and disrupts packing interactions with adjacent protein residues. Modeling

10   of the SEA mutant Y92A predicts an increase in solvent-accessible surface area for Y108 by a factor of two greater than the wild-type structure, allowing the formation of a hydrogen bond to the carboxylate group of D70 and thus disrupting key anchoring and

15   recognition points for HLA-DR1. This effect is expected to be somewhat less in SEB due to the longer side chain at E67. Substitution of SEB Y115 with alanine also resulted in 100-fold reduction of binding. In contrast, the same replacement of Y108 in

20   SEA yielded little to no change in DR1 binding (**Fig. 2a**), suggesting the primary importance of SEA residues Y92 and D70 for stabilizing interactions with K39. The K39 side chain of DRα forms a strong ion-pair interaction with the SEB E67 carboxylate group and

25   hydrogen bonds with the hydroxyl groups of Y89 and Y115. Substitution of SEB E67 by glutamine reduced binding affinity by 100-fold (**Fig. 2**), reflecting the replacement of the strong ionic bond with a weaker hydrogen bond. To optimize ion-pair interactions of

30   the analogous SEA site, the shorter carboxylate side chain of D70 is predicted to shift K39 of DRα, weakening interactions with SEA Y108. The substitution of alanine for SEA Y108 is thus more easily accommodated than the homologous substitution

35   of SEB Y115, without loss in DR1 binding.

Comparisons of the polar pocket with other bacterial superantigens were then made. SEC1-3 and SPE-A have conserved the critical DR1 binding-interface residues (**Fig. 1**), and share with SEB and

5   SEA secondary structural elements of the DR1-binding surfaces. Asparagine in SED (N70) replaces the acidic side chain present in SEA, SEB, SPE-A and SEC1-3. Accordingly, for SED the salt bridge of the polar pocket is likely to be replaced by a hydrogen bond.

10   Overall DR1 affinities for SED and SEA appeared to be equivalent (**Fig. 2b**), indicating that other interactions may compensate for the absence in SED of the ion-pair found in the other superantigens. For the case of TSST-1, mutating DRα residues K39 to

15   serine or M36 to isoleucine has been shown to greatly reduce binding [Panina-Bordignon *et al.* (1992) *J. Exp. Med.* **176**: 1779-1784]. Although primarily hydrophobic, the critical TSST-1 structural elements are conserved with the SEA and SEB polar binding

20   pocket. SEB residues Y89 and Y115 are homologous to T69 and I85 in TSST-1, respectively, and SEB E67 is replaced by I46. These TSST-1 residues are positioned in a conserved ß-barrel domain found in both SEB and SEA. However, the TSST-1 site lacks polarity

25   equivalent to SEB/SEA, and hydrogen bonding with the hydroxyl of TSST-1 residue T69 would require that DRα K39 extend 5 Å into the pocket. TSST-1 binding utilizes an alternative strategy [Kim *et al.* (1994) *Science* **266**: 1870-1874] consisting of hydrophobic

30   contacts centered around residue I46, and potential ionic or hydrogen bonds bridging DRα residues E71 and K67 to R34 and D27, respectively, of TSST-1.

The hydrophobic region of the binding interface between SEB and the HLA-DR1 molecule consists of SEB

35   residues 44-47, located in a large reverse turn

connecting ß-strands 1 and 2 of SEB. These residues appear to make strong electrostatic interactions with DRα through their backbone atoms. The mutation of L45 to an arginine reduced overall HLA-DR1 binding greater

5 than 100-fold (**Fig. 2b**), attributable to the less energetically favorable insertion of a highly charged residue into a hydrophobic depression on the DR1 molecule. The modeled DR1-SEA complex presents similar interactions with the SEA backbone atoms, with

10 the exception of a glutamine (Q49) replacing SEB Y46. Mutation of L48 to glycine in SEA (homologous to L45 of SEB) has been reported to decrease T-cell responses. SEB L45 and the comparable L30 of TSST-1 are the most extensively buried residues in the DR1

15 interface. The leucine is conserved among the bacterial superantigens (**Fig. 3**) and may provide the necessary hydrophobic structural element for surface complementarity with DR1, consistent with the mutagenesis data for SEB and SEA.

20 The inventor has performed similar structure and function studies with TSST-1, SEC1 and SPE-A.

EXAMPLE 3

Molecular modelling and structural studies of
25 staphylococcal and streptococcal superantigens: Some
interactions of bacterial superantigens with MHC class
II receptors are not conserved but are less important
than the hydrophobic loop and polar pocket binding
sites.

30 In determining the overall affinity of the superantigen for DR1, a contributory role is played by structural variations around the common binding motifs. A short, variable structured, disulfide-bonded loop is found in SEA and a homologous longer

loop in SEB. The SEB residue Y94, contained within this loop, forms hydrophobic interactions with L60 and A61 of the DRα subunit. Replacement of Y94 with alanine partially inhibits DR1 binding (**Fig. 2a,b**).

5   An alanine is found in SEA (A97) and SEE at the position equivalent to SEB Y94, and mutating this residue in SEA to tyrosine results in disrupted instead of stabilized interactions with DR1 (**Fig. 2a**). Although the disulfide loops differ in structure

10 between SEA and SEB, A97 apparently contributes to the DRα binding interface in a manner similar to Y94 of SEB. Because TSST-1 lacks a disulfide loop, similar contacts with DRa are replaced by interactions with β-strands of TSST-1. In a like manner, the absence of a

15 salt bridge between the residues K39 of DRα and E67 of SED is apparently compensated for by stabilizing interactions occurring outside of the otherwise conserved dominant binding surfaces (**Fig. 2a**).

20   **EXAMPLE 4**

Molecular modelling and structural studies of staphylococcal and streptococcal superantigens: Superantigen interactions with T-cell antigen receptors.

25   The amino acid residues in contact with TCR are located in regions of high sequence variability, presenting a unique surface for interaction with the TCR. Residues implicated in TCR interactions by mutagenesis of SEA and SEB reside in variable loop

30 regions, while TSST-1 mutants that affect TCR binding are mainly located in an α helix [Acharya, R.K. *et al.* (1994) *Nature* **367**, 94-97; Kim, J. *et al.* (1994) *Science* **266**, 1870-1874]. Specifically, mutations that diminish T-cell receptor recognition of SEB include

residues N23, Y61, and the homologous SEA N25 or Y64 **(Fig. 2c)**. SEA residues S206 and N207 also control T-cell responses [Hudson, *et al.*(1992) *J. Exp. Med.* **177**: 175-184]. Mutants of the polar binding pocket, ,

5 SEA Y92A and SEB Y89A, equivalently reduced T-cell responses **(Fig. 2c)**, reflecting the observed decreases in DR1-binding **(Fig. 2a, b)**. While supporting reduced T-cell responses, mutants SEA Y64A and SEB Y61A retained normal affinities for DR1 **(Fig.**

10 **2a-c)**.

EXAMPLE 5

Animal models for determining biological activity of bacterial superantigens: Mouse.

When compared to primates, mice are not very

15 susceptible to the toxic effects of SE, and we therefore sought to increase sensitivity with a potentiating dose of lipopolysaccharide (LPS) from Gram-negative bacteria [Stiles *et al.* (1993) *Infect. Immun.* **61**, 5333-5338]. There was no apparent effect

20 in control animals injected with any of the SE (up to 30 µg/mouse) or LPS (150 µg/mouse) alone (Table 1). Incremental injections of LPS were also not lethal, when given in amounts up to 250 µg/mouse (data not shown). However, mice died between 24-48 h after SE

25 and LPS were given to the same animal (Table 1). SEA was much more toxic than either SEB or SEC1 and the calculated LD50 (µg toxin/kg) of SEA, SEB, and SEC1 with 95% fiducial limits was 18.5 (6.5, 38.5), 789.0 (582.5, 1044.5), and 369.0 (197.5, 676.0),

30 respectively.

50

TABLE 1. Titration of SEA, SEB, and SEC₁ in the C57BL/6 mouse lethality assay

Stimulus[a]	% Lethality (no. of mice tested) with the following dose of SE, in micrograms/mouse[b]:			
	30	10	1	0.1
SEA + LPS	93(15)[b]	85(20)	80(15)	20(10)
SEB + LPS	80(15)	27(15)	0(15)	0(15)
SEC₁+ LPS	80(10)	60(10)	10(10)	0(10)

[a]LPS was injected into each mouse (150ug) 4 h after the SE injection. Control mice injected with 150 ug of LPS (n=20) or 30 ug of SEA, SEB, or SEC1 (n=10) survived.
[b]Results are from a combination of separate experiments with five mice per experiment.

The role of MHC class I and class II molecules in SE toxicity, potentiated by LPS, was addressed by using transgenic, MHC-deficient mice (Table 2). Class II-deficient animals were unaffected by a dose of SE (30 µg) plus LPS (150 µg) that was lethal for 93% of wild-type and 30% of class I-deficient mice. Mononuclear cells from class II-deficient animals were not able to present SEA, as measured by proliferative responses. MHC class I-deficient cells were functional in supporting T-cell proliferation, but at levels <30% of the proliferative response supported by MHC-wild-type presenting cells (Table 3). Cell surface expression levels were normal, when compared to nontransgenic C57BL/6, for A^b in class I-deficient mice, and K^b/D^b in class II-deficient mice. The T-cell responses of MHC class I- or class II-deficient mice were essentially equivalent to wild-type when SEA

was presented by mononuclear cells expressing both class I and II molecules (Table 3).

TABLE 2. Lethality of SEA and SEB in C57BL/6 mice lacking MHC class I or class II

Stimulus[a]	% Lethality (no. of mice tested) with the following MHC class phenotype		
	I⁻II⁺	I⁺II⁻	I⁺II⁺
SEA + LPS	30(10)	0(5)	93(15)
SEA + LPS	ND[b]	0(5)	80(15)
SEA only	0(2)	0(2)	0(2)
SEB only	ND[b]	0(2)	0(2)
LPS only	0(5)	0(5)	0(5)

[a] Mice were injected with 30 ug of SEA or SEB and, 4h later, with 150 ug of LPS, as indicated. Control mice were injected with only SEA, SEB, or LPS.
[b] ND, not determined.

Table 3. Mouse T-cell responses to SEA are MHC class II-dependent

T-cell/APC source[3]	T-cell responses[1]	
	0.1 μg/ml SEA	1 μg/ml SEA
Wild-type C57/BL6 mouse/autologous	430,000 cpm[2]	700,000 cpm
MHC class I knock-out C57/BL6 mouse/autologous	117,000 cpm	167,000 cpm
MHC class II knock-out C57/BL6 mouse/autologous	8,000 cpm	33,000 cpm
Wild-type C57/BL6 mouse/wild-type	305,000 cpm	307,000 cpm
MHC class I knock-out	420,000 cpm	445,000 cpm

C57/BL6 mouse/wild-type

MHC class II
knock-out C57/BL6   310,000 cpm   322,000 cpm
5   mouse/wild-type

[1]Cultures of mononuclear cells derived from mouse spleens,
cultured for 3 d with the indicated amount of SEA.

10   [2]Data represent the mean of triplicate determinations (<10 SEM)
of [^{3}H]thymidine incorporation.

[3]Antigen presenting cells (APC) were isolated from spleens of
the indicated mouse strain and added to cultures.
15

    The serum levels of TNFα, IL-1α, IL-6, and IFNγ
in mice injected with SEA, LPS, or SEA plus LPS were
measured at various times following injection (**Fig.
4**). Compared to mice injected with either SEA or LPS
20   alone, the serum levels of TNFα, IL-6, and IFNγ had
increased 5-, 10-, and 15-fold, respectively, in
animals given SEA plus LPS. SEA alone did not elicit
any detectable increase of TNFα, IL-6, or IFNγ above
background. In contrast to the other cytokines, IL-1α
25   levels in mice injected with SEA plus LPS resulted in
a simple additive effect.

    Serum levels of TNFα, IL-6, and IFNγ were maximal
2-4 h after the LPS injection, but returned to normal
by 10 h. The concentration of IL-1α in mice given SEA
30   plus LPS had also peaked 2 h after the LPS injection,
but stayed above background for the remaining
determinations. Levels of IL-1α in mice given only
LPS or SEA peaked at 4 and 6 h, respectively. Unlike
profiles for other cytokines, the highest amount of
35   IL-1α in mice injected with SEA and LPS corresponded
to the peak stimulated by SEA, but not LPS.

    This animal model was used in various stages of
developing the inventions, as a means of assessing the
physiological activity of mutated superantigens.

Control animals survived the maximum dose of either SE
or LPS, while mice receiving both agents died. Wild-
type SEA was 43-fold more potent than SEB and 20-fold
more potent than SEC1. By using BALB/c mice the

5   toxicity of SEB was 10-20 fold higher. These data
confirmed that the toxicity of SE was mainly exerted
through a mechanism dependent on expression of MHC
class II molecules and was linked to stimulated
cytokine release. Thus this was a relevant

10   preclinical model that could be used to predict human
responses.

EXAMPLE 6

Animal models for determining biological activity

15   of bacterial superantigens: Rhesus monkey

The physiological responses of the rhesus monkey
to bacterial superantigens is probably identical to
humans, with the exception of sensitivity [Bavari and
Ulrich (1995) *Clin. Immunol.Immunopath.* **76**:248].

20   Generally SEB intoxicated monkeys developed
gastrointestinal signs within 24 hours post-exposure.
Clinical signs were mastication, anorexia, emesis and
diarrhea. Following mild, brief, self-limiting
gastrointestinal signs, monkeys had a variable period

25   of up to 40 hours of clinical improvement. At
approximately 48 hours post-exposure, intoxicated
monkeys generally had an abrupt onset of rapidly
progressive lethargy, dyspnea, and facial pallor. If
given a lethal dose, death occurs within four hours of

30   onset of symptoms. Only SEB has been used in
challenges of rhesus monkeys to determine
physiological/pathological effects. Human responses
to bacterial superantigens are characterized by a
rapid drop in blood pressure, elevated temperature,

and multiple organ failure-the classical toxic shock syndrome (TSS). However, the respiratory route of exposure may involve some unique mechanisms. The profound hypotension characteristic of TSS is not

5   observed, and respiratory involvement is rapid, unlike TSS. Fever, prominent after aerosol exposure, is generally not observed in cases of SEB ingestion.

EXAMPLE 7

10   <u>Targeting receptor interactions to develop vaccines</u>.

The SEA mutants Y92A, with reduced DR1 binding, and Y64A, with reduced TCR interactions, and K14E with wild-type (control) activity were used to determine

15   the correct receptor to target for vaccine development. The binding of WT or mutant SEA was evaluated with the MHC class II expressing murine B-cell lymphoma cell line A20 (Table 4). The binding affinity of WT SEA to mouse MHC class II (H-2^d)

20   molecules was lower than that observed with human MHC class II expressing cells , reflecting the reduced toxicity that bacterial SAgs exert in mice. WT SEA, Y64A and K14E all had the same relative affinity to mouse MHC class II molecules. Similar to the results

25   obtained with human MHC class II molecules, the Y92A mutant exhibited substantially reduced binding to A20 cells (Table 4).

30

35

Table 4. Biological activity of superantigen vaccines

toxin	T-cell anergy[1]	MHC classII binding[2]	T-cell response
SEA wild type	++++	+++	+++
TCR attenuated Y64A	+	+++	+/-
MHC attenuated Y92A	-	+/-	+/-
Control K14E	++++	+++	+++

[1]Based on attenuation of T-cell response to wild-type SEA in mice immunized with the mutant or wild-type SEA.

[2]Binding to the mouse MHC class II+ A20 cells, measured by flow cytometry

The effect of WT SEA or site-specific SEA mutants on splenic mononuclear cells obtained from nonimmunized C57BL/6 (H-2^b) mice is summarized in Table 4. Both WT SEA and the control mutant K14E were potent T cell activators, effective at minimal concentrations of 10 to 100 pg/mL. However, T-cell responses to Y92A were reduced at least 100-fold, compared to SEA wild type, while Y64A-stimulated responses were slightly higher than Y92A. These results confirmed that attenuation of superantigen binding to either MHC class II or TCR molecules resulted in dramatically reduced mouse T-cell proliferation. These results may indicate that the altered toxin may compete with wild type toxin for TCR binding.

SEA WT (10 LD50), site-specific SEA mutants (10 μg/mouse each) or LPS (150 μg/mice) injected alone were nonlethal to mice (Table 5). However, combining

LPS with either WT SEA or mutant K14E resulted in 100%
lethality. For those mice receiving both LPS and WT
or K14E SEA, 80% were dead by 24 h and 100% by 48 h.
In contrast, 100% of Y92A and 80% of Y64A injected

5   mice (coadministered with LPS) survived. The average
time to death for the 20% of mice that did not
survive Y64A injection occurred at 48 to 72 h. These
in vivo data correlated well with the results obtained
with the lymphocyte cultures. It was concluded that

10 the observed attenuation of toxicity in mice was a
direct result of the reduced T-cell proliferation.

Table 5. Biologic effect of wild type (WT)
staphylococcal enterotoxin A (SEA) and SEA mutants.

15

Protein	No. live/total
WT	0/10
K14E	0/10
Y64A	8/10
Y92A	10/10

NOTE. Mice were given 10 LD_{50} (10ug) of WT or mutant SEA.
Lipopolysaccharide (150 ug/mouse) was injected 3 h later.

25

Having established that attenuation of receptor
binding resulted in reduced toxicity, we next examined
the immunogenicity of the SEA mutants. Mice were
immunized with WT or mutant SEA. Control mice

30 received adjuvant only or were left untreated. One
week before challenge with WT SEA, mice were bled and
serum antibody titers were determined for each group
(Table 6). Mice immunized with the 2 μg of Y64A or
Y92A had serum antibody titers of 1:5000 and 1:1000,

35 respectively. Immunization with 2 μg of WT SEA or

control mutant resulted in titers of 1:5,000 and
1:10,000, respectively. The highest immunizing dose
(10 µg/mouse) was most effective for all animals,
resulting in antibody titers which were greater than
5 1:10,000. All mice were challenged with 10 LD50 of WT
SEA (potentiated with LPS). The survival data
correlated well with the levels of serum antibodies in
immunized mice. All mice that were vaccinated with 10
µg of Y64A or Y92A, survived the lethal challenge dose
10 of WT SEA. Slightly less protection was afforded by
the lower vaccination dose of mutant Y64A or Y92A.
All mice immunized with both doses of WT SEA survived
the lethal challenge with WT potentiated with LPS.
Mice immunized with mutant K14E exhibited survivals of
15 100% and 80% for high and low vaccination doses,
respectively. All nonimmunized or control mice that
were vaccinated with adjuvant alone died when
challenged with WT SEA and a potentiating dose of LPS.

20 Table 6. Mice immunized with attenuated forms of
staphylococcal enterotoxin A (SEA) produce high titers
of neutralizing antibody.

Immunizing agent	Dose (ug/mouse)	Anti-SEA antibody titer*	No. live/total
WT	2	10,000-50,000	10/10
	10	10,000-50,000	10/10
K14E	2	5,000-10,000	8/10
	10	10,000-50,000	10/10
Y64A	2	5,000-10,000	6/10
	10	10,000-50,000	10/10
Y92A	2	1,000-5,000	2/10
	10	10,000-50,000	10/10
Adjuvant		50-100	0/10

NOTE. Mice were given 10 LD_{50} of wild type (WT) SEA challenge followed by potentiating dose of lipopolysaccharide (150 ug/mouse) 3 h later.

*Reciprocal of serum dilution resulting in optical
5   density reading four times above negative controls (wells containing either no SEA or no primary antibody).

EXAMPLE 8

Immune recognition of SAg mutants.

10   Bacterial SAgs induce clonal anergy of specific subsets of T cells in mice. It was possible that the loss of sensitivity to WT SEA among the mice vaccinated with the attenuated mutant forms represented a state of specific non-responsiveness
15   instead of specific immunity. To address this issue, lymphocyte responses to SEA WT were measured with splenic mononuclear cells collected 2 weeks after the third immunization. As expected, lymphocytes from mice that were immunized with WT SEA or control SEA
20   mutant showed little to no proliferation when incubated with the WT SAg. In contrast, lymphocytes obtained from control mice or those immunized with either Y64A or Y92A all responded vigorously to the WT SEA (**Fig. 5**). The TCRs used by T cells from the SEA-
25   vaccinated mice were then characterized by flow cytometry. T cells from immunized or control mice were incubated with WT SEA in culture for 7 days, followed by a 5 day expansion in IL-2 containing medium. Distinct populations of activated TCR Vß11
30   positive cells were observed with T cells from mice immunized with Y92A and Y64A, representing 48% and 40% of T cells, respectively. However, Vß11 expressing cells obtained from SEA WT or K14E immunized mice were about 1% and 6% of the total T-cell population,
35   respectively, suggesting that this subset was

nonresponsive to restimulation with the WT SAg. T cells bearing Vß 17a, 3, 7, and 10b were unchanged for all mice. It was apparent that T-cell responses to both the TCR and MHC class II binding-attenuated SEA

5   mutants were similar to each other, but differed from responses to control or WT molecules. These results suggested that an alternative, perhaps conventional antigen processing mechanism was functioning in presentation of the SAg mutants Y64A and Y92A.

10

EXAMPLE 9

Rhesus monkey immunizations with monovalent

vaccines.   (SEQ ID NO:2)

The SEA vaccine L48R, Y89A, D70R (A489270), and
(SEQ ID NO: 9)
15   SEB vaccine Y89A, Y94A, L45R (B899445), were used to immunize rhesus monkeys. The animals received a total of three i.m. injections (10-20 μg/animal), given at monthly intervals. Rhesus monkeys that were injected with these vaccines had no detectable increase of

20   serum cytokines and no apparent toxicity. The serological response of animals vaccinated with three doses of formalin-treated SEB toxoid (100 μg/injection) gave results comparable to one or two injections with B899445 (Table 7), suggesting that the

25   recombinant vaccines were very immunogenic. Immunized rhesus monkeys survived a lethal challenge with >10 LD50 of wild-type SEB (Table 7, 8). Collectively, these results suggest that the engineered SEB vaccine is safe, highly antigenic and effective at protecting

30   the immunized individual from lethal aerosol exposure to SEB.

35

Table 7. Rhesus monkey antibody responses to vaccine B899445; One injection of B899445 outperforms three injections of SEB toxoid

Vaccine[1]/animal #	Antibody response[2]	%Inhibition of T-cell response[3]	Survival SEB >20 x LD50 challenge[4]
preimmune sera /pooled	0.161	5	dead
toxoid/1	0.839	0	dead
toxoid/2	0.893	34	live
toxoid/3	1.308	57	live
toxoid/4	1.447	55	live
B899445/1	1.788	69	live
B899445/2	0.78	49	live

[1]Rhesus monkeys were immunized with one dose (20 µg injection) of B899445 vaccine or three doses of formalin-treated SEB toxoid (100 µg/injection) one month apart; both used Alum adjuvants.

[2]Sera were collected one month after the final injection. Antibody responses were determined by ELISA and the results are shown as mean optical densities of triplicate wells (± SEM).

[3]Rhesus monkey T cells, obtained from an untreated animal, were preincubated with diluted (1:70) serum from immunized monkeys and then cultured with wild type SEB. Data are shown as % of T cell responses, where serum of rhesus monkey injected with adjuvant only represented the 100% of response to wild type SEB.

[4]Rhesus monkeys were challenged by aerosol exposure and monitered for four days.

Table 8. Engineered staphylococcal entero-toxin B vaccine efficacy in rhesus monkeys

Treatment[1]	Antibody titer[2]	Immune protection[3]
Vaccine with adjuvant	>10,000	100%
Adjuvant only	<50	0%

[1]Rhesus monkeys (n=10) were injected i.m. with 10 µg of SEB vaccine with Alhydrogel adjuvant. A total of 3 immunizations, 1 month apart were given. Controls (n=2) received only Alhydrogel.

[2]Serum dilution resulting in optical density readings of four times above the negative control, consisting of no SEB or serum added to the wells.

[3]Immunized and control rhesus monkeys were challenged with >10 LD50 of wild-type staphylococcal enterotoxin B as an aerosol.

Serum from monkeys that were immunized with the genetically attenuated vaccine inhibited T-lymphocyte responses to wild type SEB (Table 7) similarly or better than monkeys that received the SEB toxoid. Collectively, these results suggest that the recombinant SAg vaccines are safe, highly antigenic, and induce protective immunity.

Serum from B899445 immunized rhesus monkeys blocked human lymphocyte responses to wild-type superantigen when tested in *ex vivo* cultures (Table 7). These data again showed that the second and third injections of vaccine were approximately equivalent in stimulating neutralizing antibody responses. Normal T-cell responses to several superantigens, including the wild-type protein, were observed in immunized animals, indicating that no specific or generalized anergy occurred (**Fig. 6**).

EXAMPLE 10

A. Multivalent superantigen vaccines: Rhesus monkey immunizations.

Rhesus monkeys were immunized with a combined
5   vaccine consisting of B899445 and A489270. Following
the third injection, antibody recognition of wild-type
bacterial superantigens was examined (**Fig.7**). High
titers of anti-SEB, SEC1 and SEA antibodies were
evident.
10

B. Mouse immunizations.

Mice (BALB/c) were immunized with a combined
vaccine consisting of SEA, SEB, SEC1 and TSST-1 (all
wild-type). The antibody responses against each
15   individual superantigen were assessed (Table 9).
Antibodies were induced against each of the component
antigens, providing sufficient levels to protect the
mice from a lethal challenge of superantigen,
potentiated with LPS. Although not shown in the
20   Table, antibody responses against SPE-A were also
observed. Mice were also immunized with individual
superantigens and antibody responses against other
superantigens were measured (Table 10). Each
individual immunogen induced partial or complete
25   protective antibody responses against all other
superantigens tested.

30

TABLE 9. Superantigen cross-reactivity of antibodies from mice immunized with individual bacterial superantigens

Immunizing[1] Toxin	Challenging[2] Toxin	ELISA[3] Titer	Neutralizing[4] Antibody
SEA	SEA	>1/25,000	100%
SEA	SEB	>1/25,000	100%
SEA	SEC1	>1/25,000	100%
SEA	TSST1	>1/10,000	100%
SEB	SEB	>1/25,000	100%
SEB	SEA	>1/10,000	100%
SEB	SEC1	>1/2,500	100%
SEB	TSST1	>1/10,000	100%
SEC1	SEC1	>1/10,000	100%
SEC1	SEA	>1/10,000	100%
SEC1	SEB	>1/25,000	100%
SEC1	TSST1	>1/10,000	100%
TSST1	TSST1	<1/10,000	100%
TSST1	SEA	<1/1,000	50%
TSST1	SEB	<1/1,000	40%
TSST1	SEC1	<1/1,000	40%

[1]Three injections with 20 ug of antigen (BALB/c mice).

[2]LPS-potentiated challenge with 10 LD_{50}s of superantigen.

[3]ELISA antibody response against an individual superantigen.

[4]Percent mice surviving an LPS-potentiated challenge (n=10).

Table 10. Multivalent superantigen vaccine. Mouse immune responses.

Immunizing toxin[1]		Challenging toxin[2]	Antibody Titer[3]	% survival
SE-A, B, C1, TSST-1		all	N/A	100%
"	"	SEA	>25,000	100%
"	"	SEB	>25,000	100%
"	"	SEC1	>25,000	100%
"	"	TSST-1	>6,400	100%

[1]Total of three injections, two weeks apart, in RIBI adjuvant.

[2]>10 X LD50, potentiated with *E. coli* lipopolysaccharide.

[3]Measured by ELISA.

EXAMPLE 11

Design of altered TSST-1 toxin vaccine, TST30.

A comprehensive study of the relationships of TSST-1 protein structure to receptor binding were undertaken to provide insight into the design of the vaccine TST30. We have discovered that TSST-1 interactions with the human MHC class II receptor, HLA-DR, are relatively weak and can be disrupted by altering only a single critical amino acid residue of the toxin. Site-directed mutagenesis of a gene encoding the toxin and expression of the new protein product in *E. coli* were then used to test the design of the vaccine. The TSST-1 gene used was contained within a fragment of DNA isolated by BglI restriction enzyme digestion of the gene isolated from a toxigenic strain of *Staphylococcus aureus* (AB259; Kreiswirth and Novick (1987) *Mol. Gen. Genet.* **208**, 84-87). The sequence of this gene is identical to all currently

known TSST-1 isolates of human origin. The wild-type TSST-1 gene can be readily cloned from a number of clinical S. aureus isolates. The DNA fragment containing the TSST-1 gene was isolated by agarose gel

5 electrophoresis and ligated inot the prokaryotic expression vector pSE380 (Invitrogen Corp.). The DNA clone consisted of sequences encoding the leader peptide and the full length of the mature TSST-1 protein *(SEQ ID NO:11)*. This engineered vaccine is currently being

10 evaluated to determine mouse and human T-cell reactivities in vitro, and lethality in mice. The TST30 vaccine consists of the following mutation introduced into the toxin molecule: leucine at amino acid residue 30 changed to arginine. Two other

15 mutations, namely Asp27 to Ala and Ile46 to Ala have also been designed. The final vaccine may incorporate one or both of these additional mutations.

    The binding interface between TSST-1 and HLA-DR consists of a large relatively flat surface located in

20 the N-terminal domain. Leucine 30 protrudes from a reverse turn on the surface of TSST-1 and forms the major hydrophobic contact with the HLA-DR receptor molecule. Mutation of the single residue leucine 30 in TSST-1 to the charged amino acid side chain of

25 arginine is predicted to disrupt this major contact with the receptor molecule, resulting in a significant reduction in DR1 binding. This mutant molecule should therefore have lost the toxin attributes of the wild-type molecule.

30 TST30 *(SEQ ID NO:12)* was expressed as a recombinant protein in E.coli, as either a periplasmically secreted protein or as a cytoplasmic product. Purification was achieved by immunoaffinity chromatography or preparative isoelectric focusing after an initial ion-

35 exchange CM-Sepharose enrichment step. The method of

purification was not critical to the performance of the vaccine. Lipopolysaccharide contaminants, resulting from expression in a Gram-negative bacterium, were readily removed (as determined by

5 limulus assay) using a variety of standard methods. The final purified vaccine is not toxic to mice at levels equivalent to 10 LD_{50} of the native TSST-1. No indicators of toxicity were found in surrogate assays of human T-cell stimulation.

10 Conclusive vaccine studies demonstrating that TST30 is highly antigenic and induces protective immunity are in progress in a mouse animal model. Mouse lethality is achieved at less than 1 ug/animal when a potentiating signal like lipopolysaccharide

15 from Gram-negative bacteria (LPS) is provided. When coadministered with LPS, wild-type TSST-1 is 100% lethal to mice (10 LD_{50}). Mice receive three injections (two weeks between injections) of 20 ug/mouse in alhydrogel and protection against the

20 lethal effects of 10 LD_{50} of TSST-1 are assessed.

EXAMPLE   12

Design of altered SPEA toxin vaccine, SPEa42

The SPEa interactions with human MHC class II

25 receptor, HLA-DR, are relatively weak and can be disrupted by altering only a single critical amino acid residue of the toxin. Site-directed mutagenesis of a gene encoding the toxin and expression of the new protein product in E.coli were then used to test the

30 design of the vaccine. The SPEa gene used was clone from a SPEa-toxigenic strain of Streptococcus by using specific DNA oligonucleotide primers and the polymerase chain reaction method. The sequence of this gene is identicla to SPEa isolates of human

35 origin known within the public domain. The DNA

fragment containing the SPEa gene was isolated by agarose gel electrophoresis and ligated into a prokaryotic expression vector (pETx or pSE380). The DNA clone consisted of sequences encoding the leader

5 peptide and the full length of the mature SPEa protein or SPEa42 *(SEQ ID NO:15)* without a leader sequence. We recognize that there are additional ways to express or produce the mature SPEa vaccine. The SPEa vaccine consists of the following mutation introduced into the toxin

10 molecule: leucine at amino acid residue 42 changed to arginine.

The binding interface between SPEa and HLA-DR is predicted to consist of contacts located in the N-terminal domain that are conserved with other

15 bacterial superantigens. Leucine 42 of SPEa is predicted to protrude from a reverse turn on the surface of SPEa and form a major hydrophobic contact with the HLA-DR receptor molecule. Mutation of the single residue leucine 42 in SPEa to the charged amino

20 acid side chain of arginine is predicted to disrupt this major contact with the receptor molecule, resulting in a significant reduction in DR1 binding. This mutant molecule should therefore have lost the toxin attributes of the wild-type molecule.

25 SPEa42 *(SEQ ID NO:16)* was expressed as a recombinant protein in E.coli, as either a periplasmically secreted protein or as a cytoplasmic product. Purification was achieved by immunoaffinity chromatography or preparative isoelectric focusing after an initial ion-

30 exchange CM-Sepharose enrichment step. The method of purification was not critical to the performance of the vaccine. Lipopolysaccharide contaminants, resulting from expression in a Gram-negative bacterium, were readily removed (as determined by

35 limulus assay) using a variety of standard methods.

The final purified vaccine is not toxic to mice at levels equivalent to 10 LD_{50} of the native TSST-1. No indicators of toxicity were found in surrogate assays of human T-cell stimulation.

5   Conclusive vaccine studies demonstrating that SPEa42 is highly antigenic and induces protective immunity are in progress in a mouse animal model. Mouse lethality is achieved at less than 1 ug/animal when a potentiating signal like lipopolysaccharide

10   from Gram-negative bacteria (LPS) is provided. When coadministered with LPS, wild-type SPEa is 100% lethal to mice (10 LD_{50}). Mice receive three injections (two weeks between injections) of 20 ug/mouse in alhydrogel and protection against the lethal effects of 10 LD_{50} of

15   SPEa are assessed

EXAMPLE   13

Design of altered superantigen toxin vaccine,

SEC45

20   For Staphylococcal enterotoxin C1 (SEC1), the (SEC ID NO: 14) leucine at position 45 was changed to lysine (SEC45)$_A$. This mutation is anticipated to prevent SEC1 from interacting with the MHC class II receptor by sterically blocking the hydrophobic loop (centered

25   around leucine 45) from binding to the alpha chain of the receptor. SEC1 is more closely homologous to SEB than SEA or the other superantigen toxins. The presence of zinc in SEC1 may impart additional binding characteristics that allow, in some cases, this

30   superantigen toxin to bind to T-cell antigen receptors without the required MHC class II molecule interactions. To circumvent the binding to T-cell antigen receptors, mutations of SEC1 residues N23

(changed to alanine), V91 (changed to lysine) are being performed.

5

10

15

20

25

30